

Modeling Cell Signaling

Bill Hlavacek

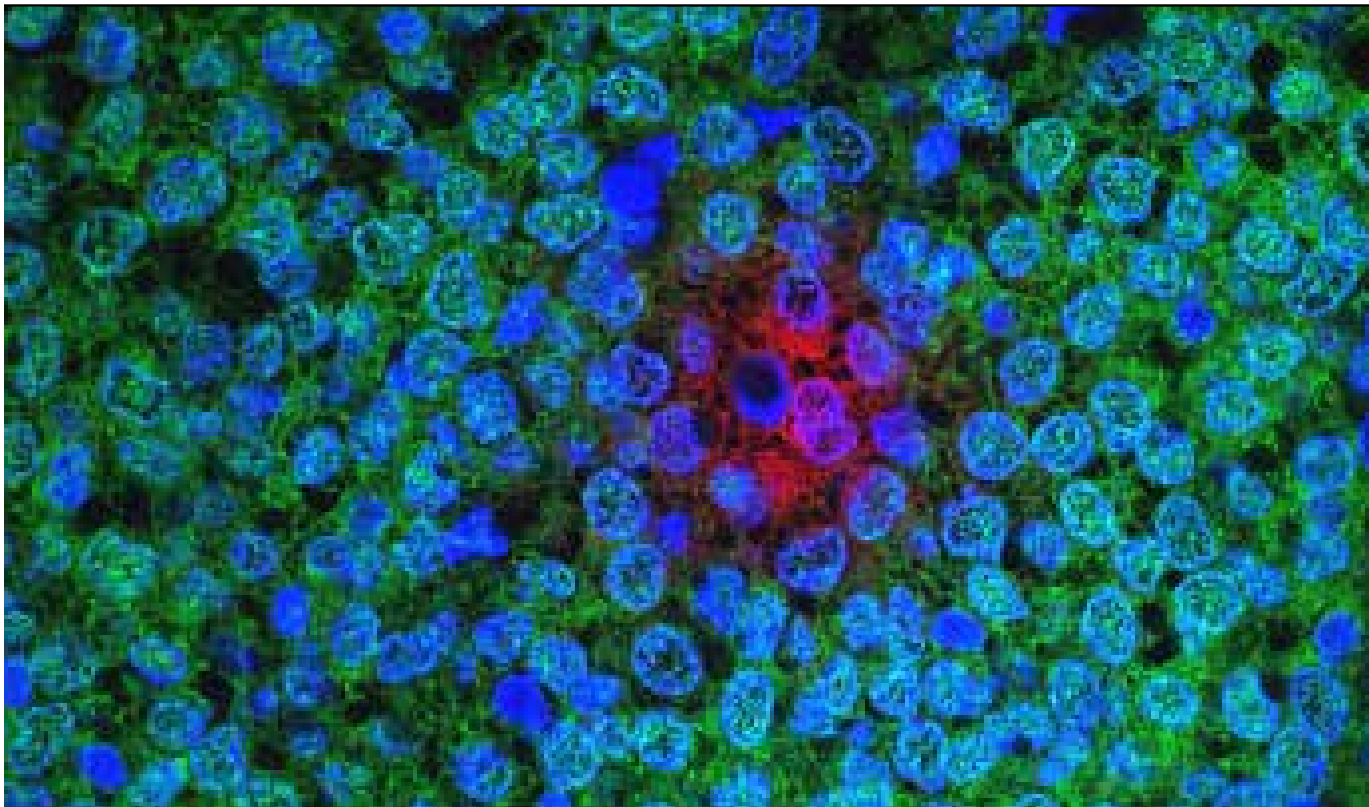
Theoretical Biology & Biophysics Group

Theoretical Division

Brian Munsky, William S. Hlavacek, and Lev S. Tsimring, editors

QUANTITATIVE BIOLOGY

Theory, Computational Methods, and Models



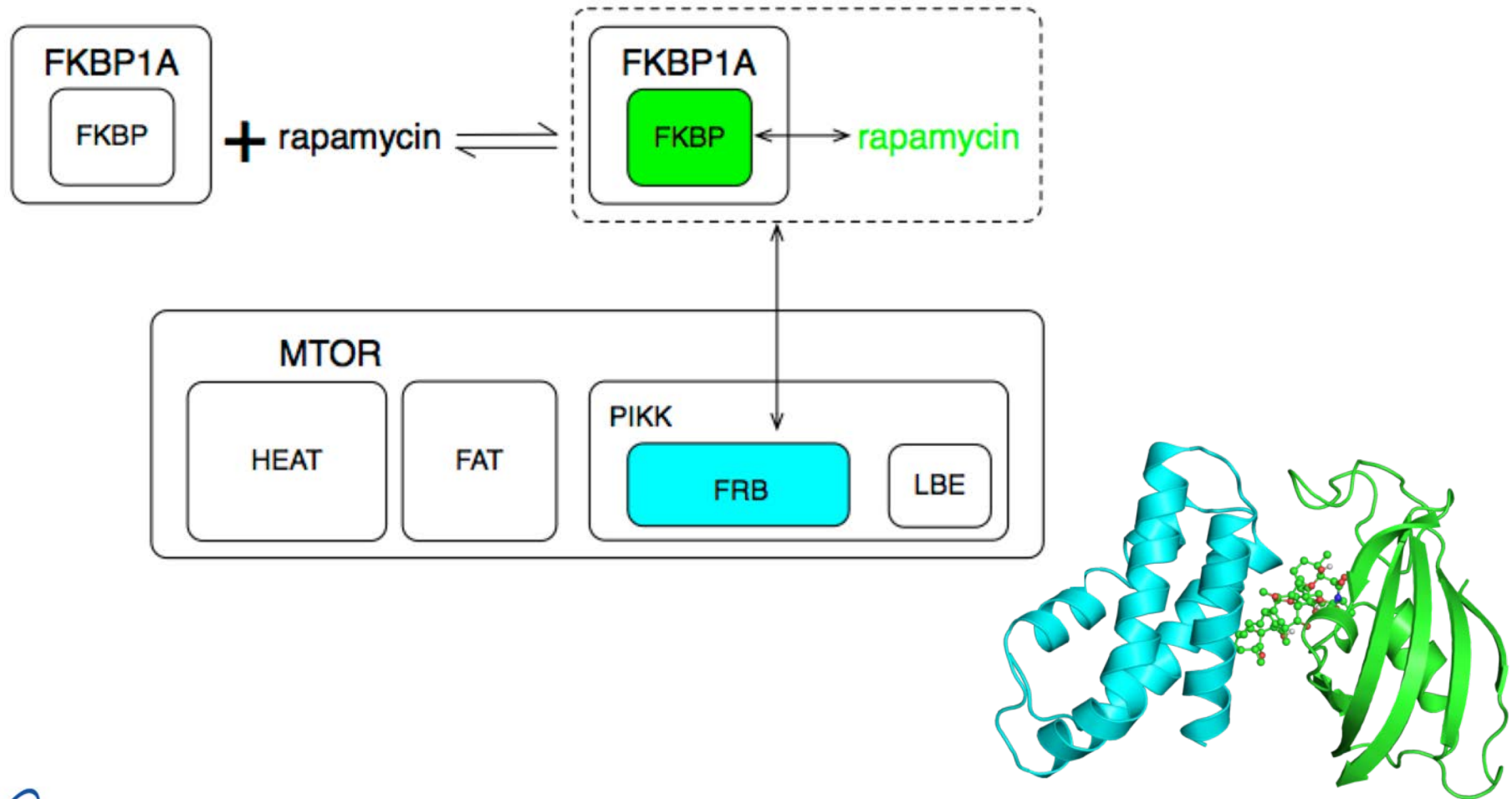
Value added by modeling of cellular regulatory systems

- **We can use models to organize and evaluate information**
 - To think with greater rigor and precision
 - To discover knowledge gaps
 - To identify key quantitative factors that affect system behavior
 - To summarize observations and preserve knowledge
- **We can analyze models to obtain insights and generate hypotheses**
 - To elucidate general design principles
 - To explain counterintuitive behavior
 - To enhance experimental efforts (e.g., through experimental design)
 - To guide interventions

Influences within the AMPK-MTORC1-ULK1 network



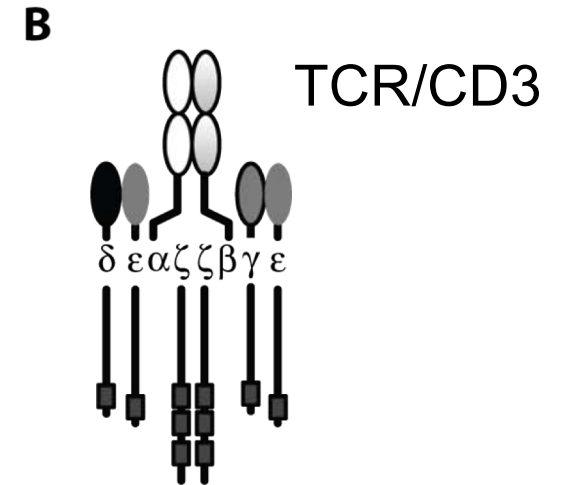
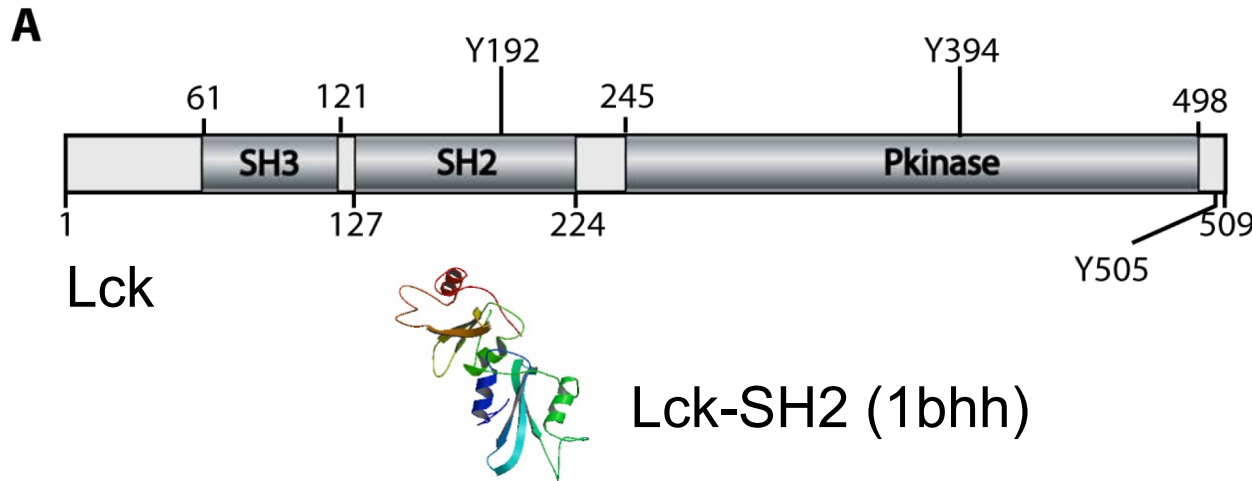
Illustration of details involved in tracking site dynamics



Outline

1. **Features of signaling proteins**
2. Combinatorial complexity: the key problem solved by rule-based modeling
3. Basic concepts of rule-based representation of biomolecular interactions
4. Simulation methods for rule-based models (indirect and direct)
5. Exercises (computer lab)

A signaling protein is typically composed of multiple components (subunits, domains, and/or linear motifs) that mediate interactions with other proteins

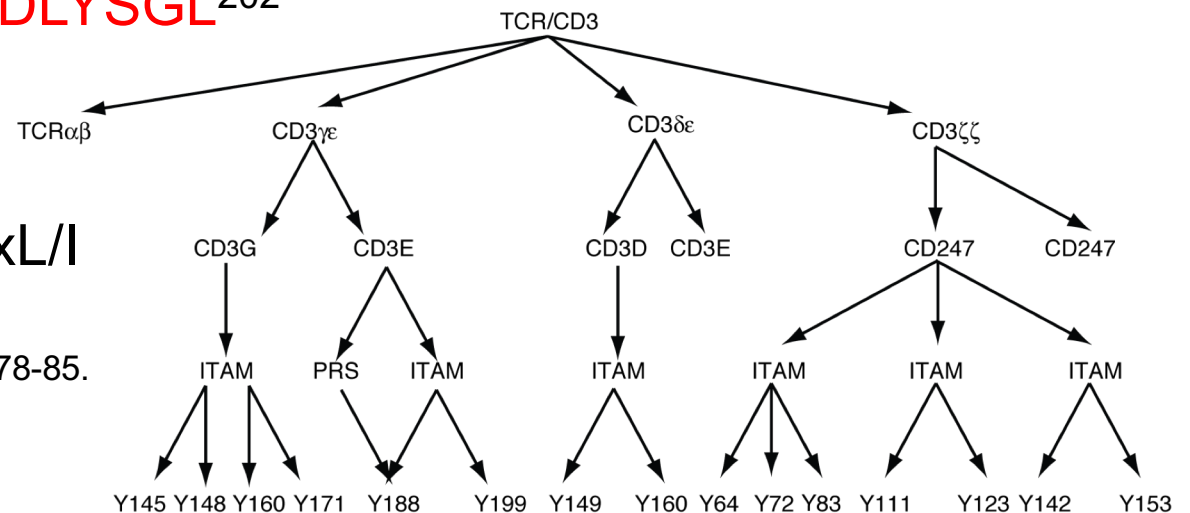


CD3E: 184PNPDYEP^{ITAM}IRKGRDLYSGL^{ITAM}202

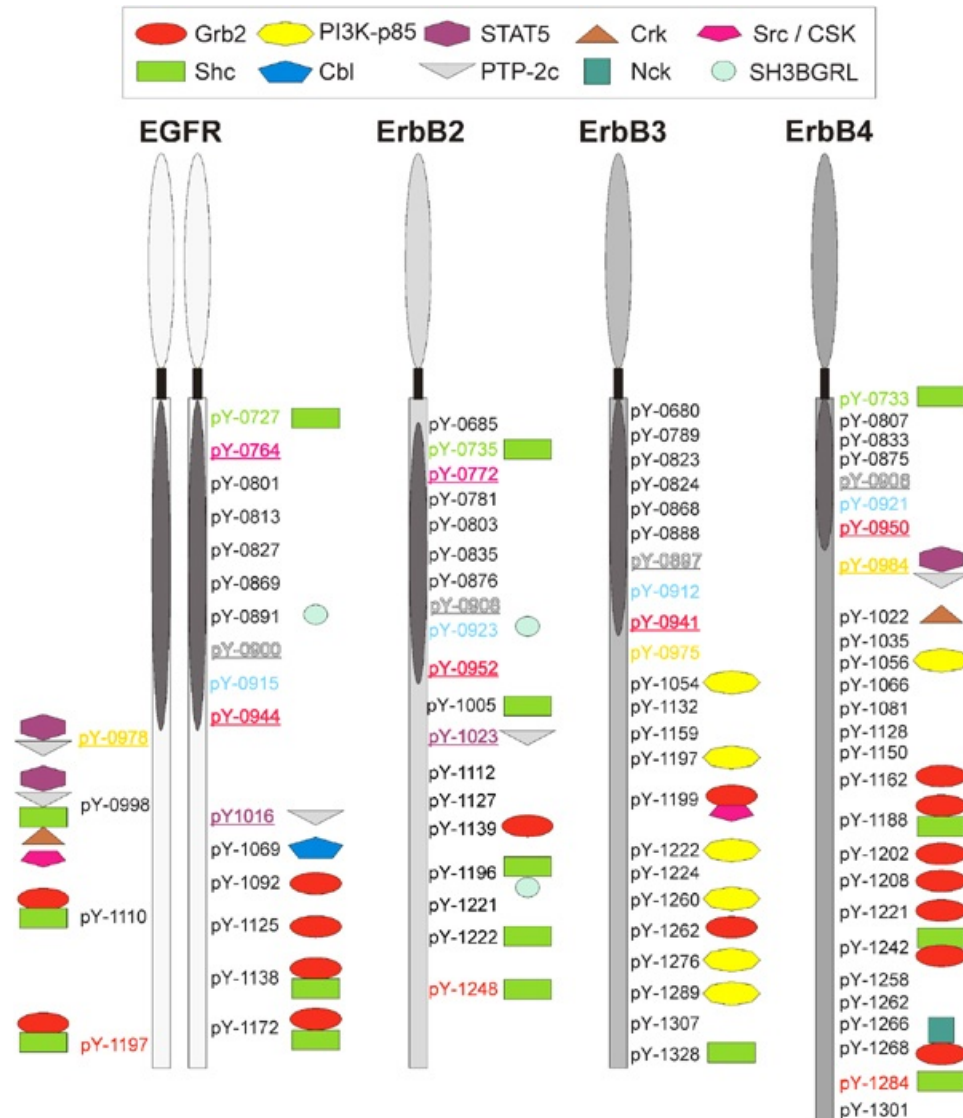
PRS: PxxDY

ITAM: YxxL/I(x₆₋₈)YxxL/I

Kesti T et al. (2007) J. Immunol. 179:878-85.



Domain-motif interactions are often controlled by post-translational modifications



Schulze WX et al. (2005)
Mol. Syst. Biol.

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Complexity arises from post-translational modifications

Epidermal growth factor receptor (EGFR)

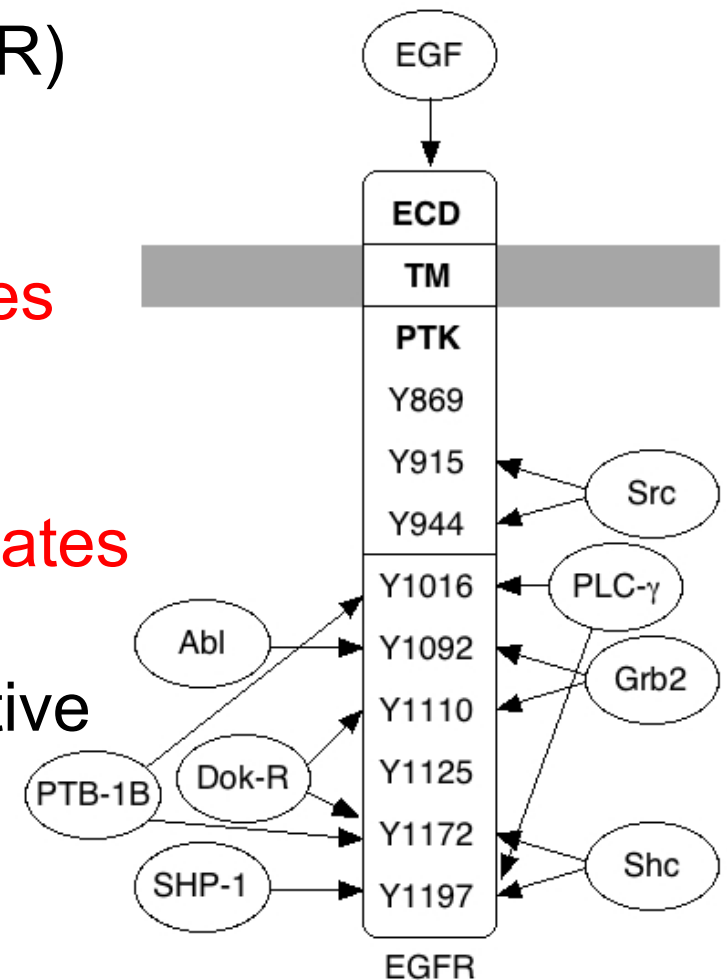
9 sites => $2^9=512$ phosphorylation states

Each site has ≥ 1 binding partner

=> more than $3^9=19,683$ total states

EGFR must form *dimers* to become active

=> more than 1.9×10^8 states

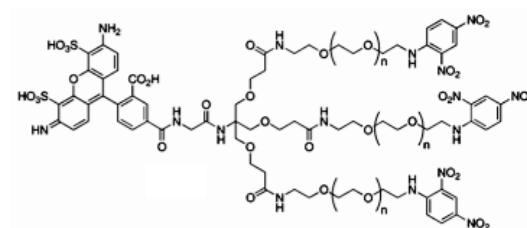


Complexity arises from oligomerization/aggregation

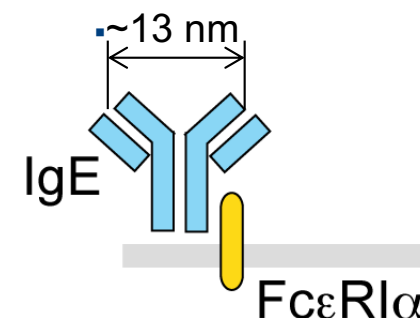
Mahajan et al. (2014) *ACS Chem Biol* **9**: 1508-1519.



DF3

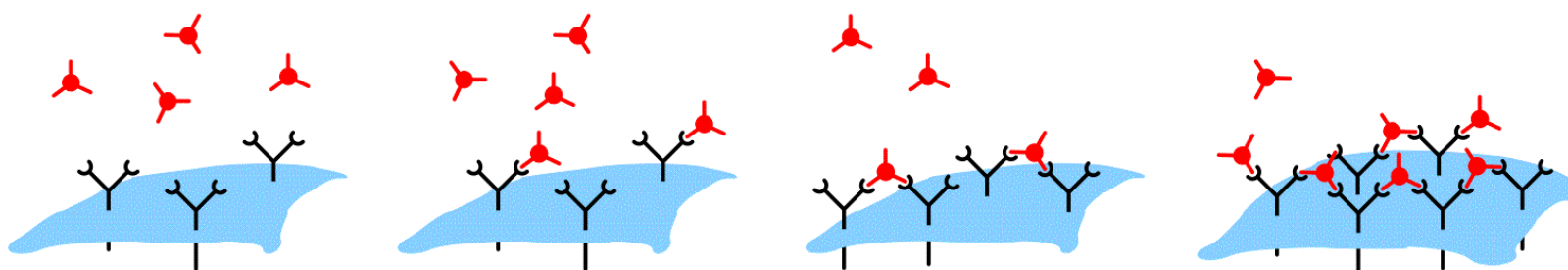


Compound 6a



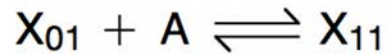
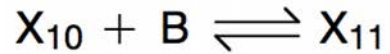
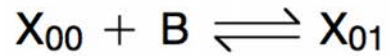
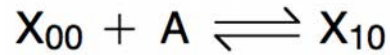
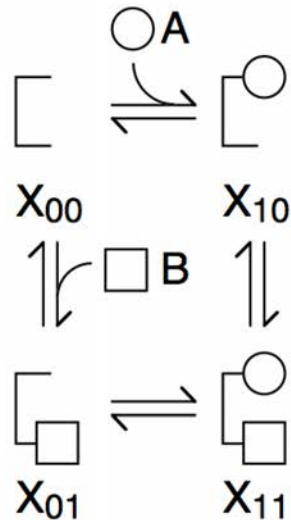
Ara h 1 (major peanut allergen), PDB 3S7E

Posner et al. (2007) *Org Lett* **9**: 3551



The textbook approach

Conventional representation of a biochemical reaction network



6 chemical species

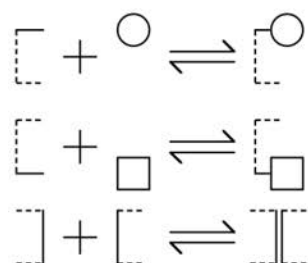
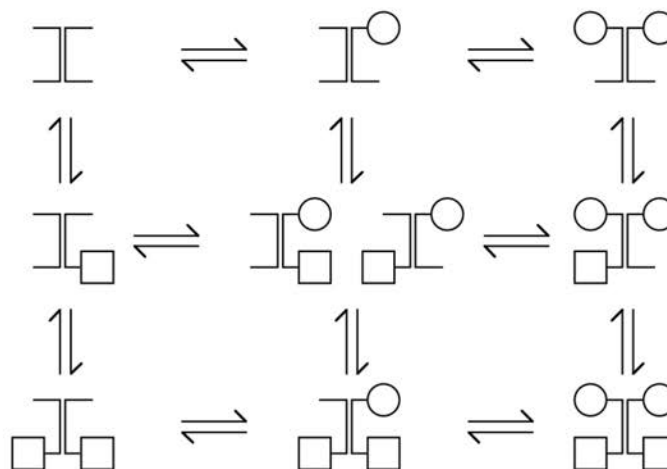
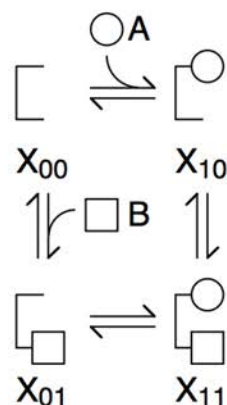
8 unidirectional reactions

$$d[X_{00}]/dt = -k_{+1}[A][X_{00}] + k_{-1}[X_{10}] - k_{+2}[B][X_{00}] + k_{-2}[X_{01}]$$

Network (model) size tends to grow nonlinearly (exponentially) with the number of molecular interactions

Network size increases nonlinearly when an extra interaction is considered

16 chemical species
60 unidirectional reactions



There are only three interactions. We can use a “rule” to model each of these interactions.

Science's STKE re6 (2006)

Rule-based modeling solves the problem of combinatorial complexity

■ Inside a Chemical Plant

- Large numbers of molecules...
- ...of a few types
- Conventional modeling works fine (a good idea since Harcourt and Esson, 1865)

■ Inside a Cell

- Possibly small numbers of molecules...
- ...of many possible types
- Rule-based modeling is designed to deal with this situation (new)

Outline

1. Features of signaling proteins
2. Combinatorial complexity: the key problem solved by rule-based modeling
3. **Basic concepts of rule-based representation of biomolecular interactions**
4. Simulation methods for rule-based models (indirect and direct)
5. Exercises (computer lab)

Rule-based modeling: basic concepts

Graphs represent molecules/complexes, their component parts, and “internal states”

collections of same-colored vertices represent “molecule types”

vertices represent “sites”

vertex labels represent “states”

edges represent bonds

connected molecule types represent complexes

Graph-rewriting rules represent molecular interactions

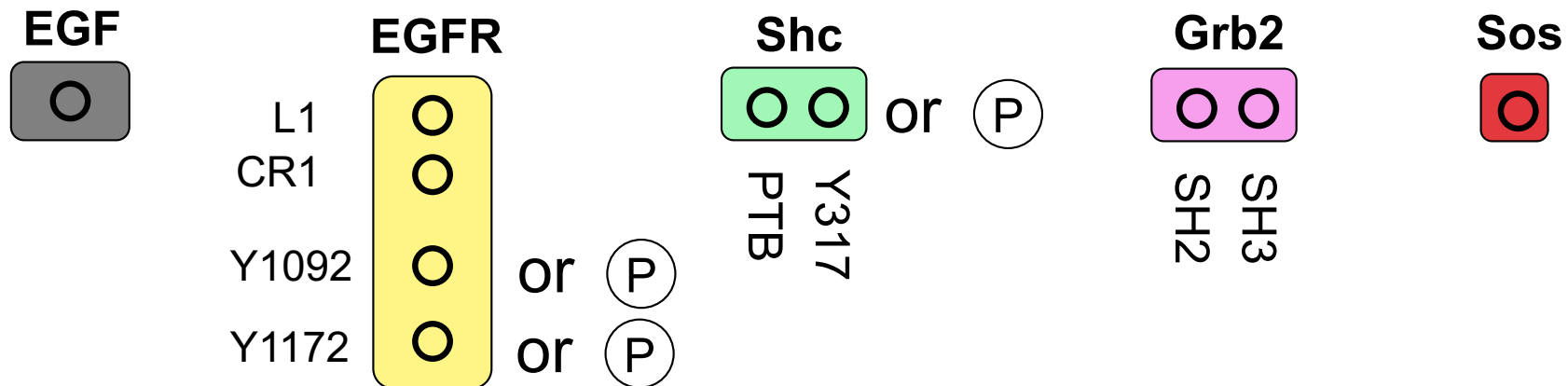
addition of an edge to represent bonding

removal of an edge to represent dissociation

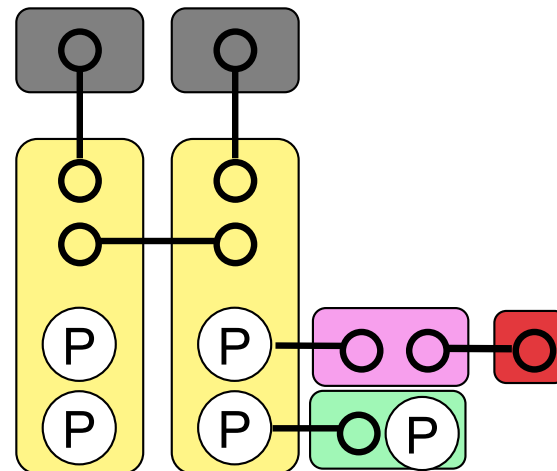
change of a vertex label to represent change of state

(e.g., change of conformation, location, or PTM status)

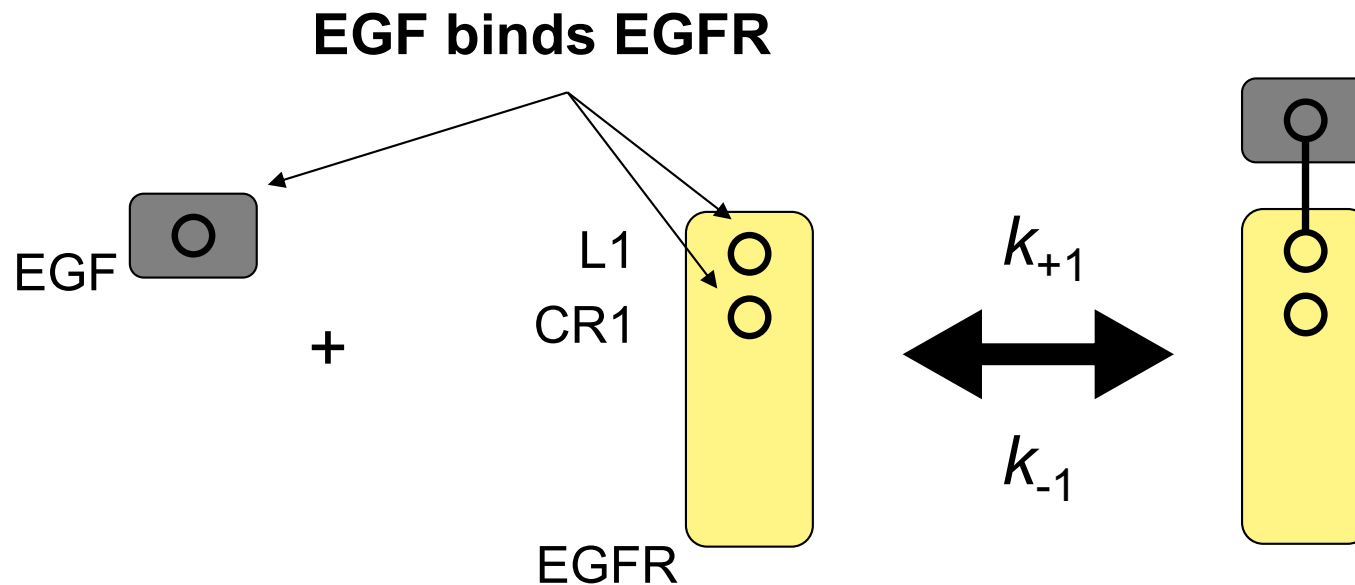
The site graphs of a model for EGFR signaling



No need to introduce a unique name (e.g., X_{123} or ShP-RP-G-Sos) for each chemical species, as in conventional modeling



A rule for EGF-EGFR binding



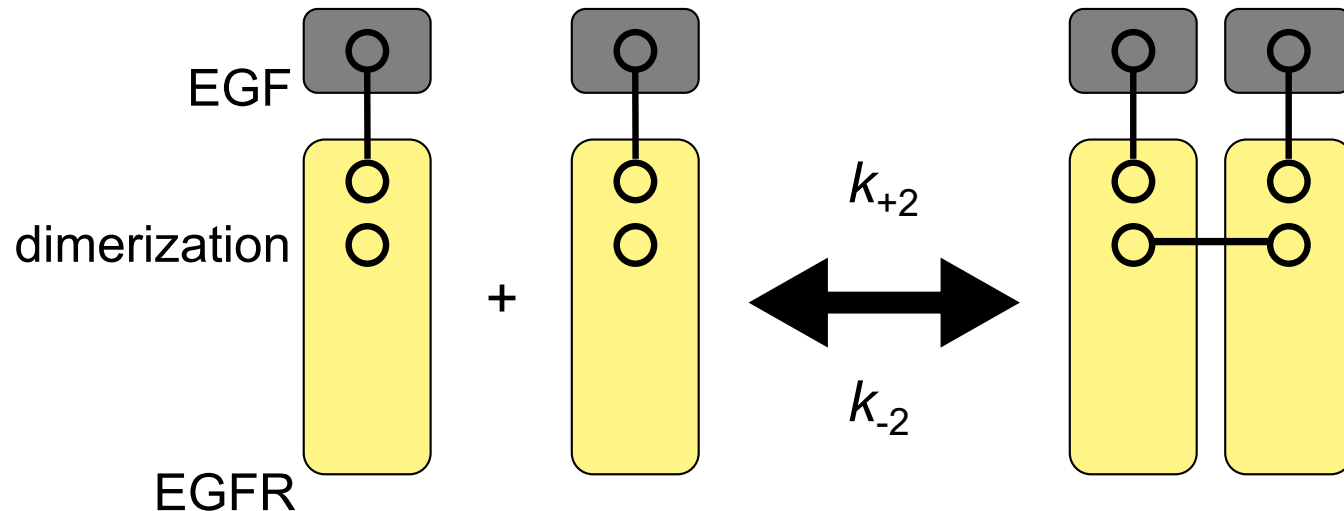
begin reaction rules



end reaction rules

A rule for ligand-dependent EGFR dimerization

EGFR dimerizes (600 reactions are implied by this one rule)



No free lunch: According to this rule, dimers form and break up with the same fundamental rate constants regardless of the states of cytoplasmic domains, which is a simplification.

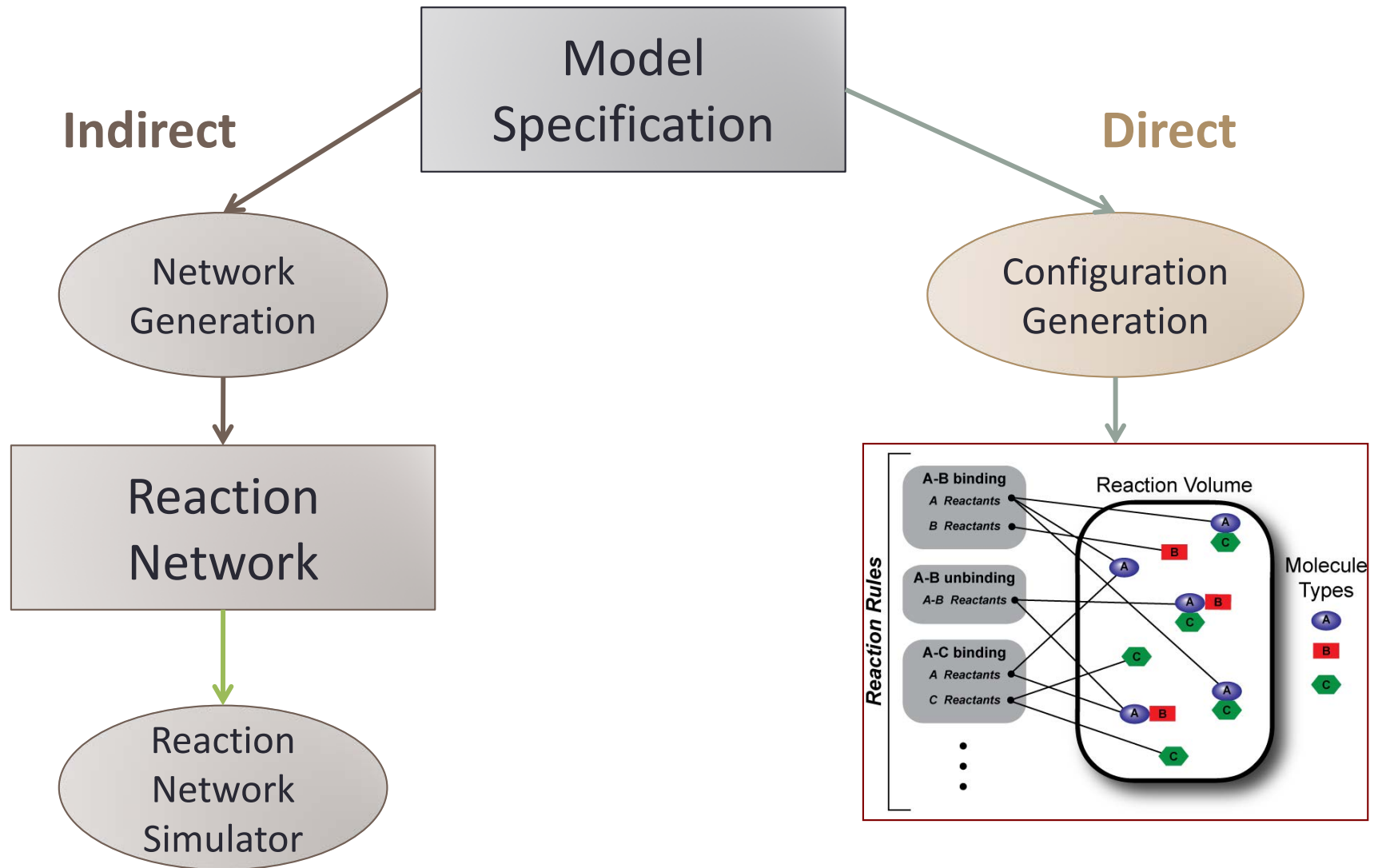
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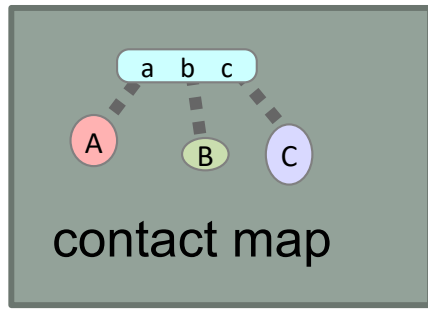
Many different traditional simulation techniques are compatible with RBM

1. Ordinary differential equations (ODEs) - **BioNetGen**
 - One equation per chemical species in the reaction network
 - Each reaction contributes a negative term to a reactant's equation and a positive term to a product's equation
2. Markov chains – BioNetGen + **NFsim**
 - Gillespie's method or stochastic simulation algorithm (SSA) or KMC
 - Each trajectory represents one sample from probability space of the chemical master equation (CME)
3. Partial differential equations (PDEs) - **VCell**
 - Species concentrations are resolved in space
4. Particle-based stochastic spatial simulations – **Smoldyn** + **MCell**
5. Force field- or potential-based calculations with excluded volume and orientation constraints (molecular dynamics) - **SRSim**

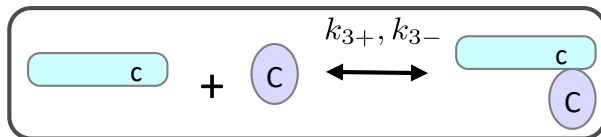
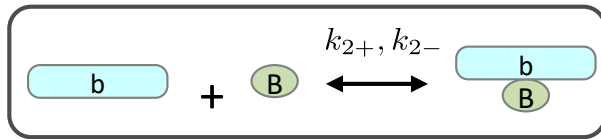
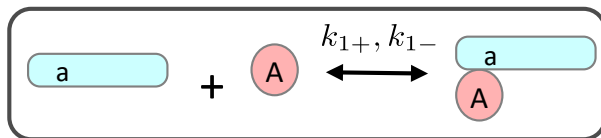
Two types of methods for simulating RBMs



Indirect Methods – Network Generation



reaction rules



seed species

S, A, B, C

S	+	A	$\xrightleftharpoons[k_{1-}]{k_{1+}}$	SA	1
SB	+	A	$\xrightleftharpoons[k_{1-}]{k_{1+}}$	SAB	2
SC	+	A	$\xrightleftharpoons[k_{1-}]{k_{1+}}$	SAC	3
SBC	+	A	$\xrightleftharpoons[k_{1-}]{k_{1+}}$	SABC	4

S	+	B	$\xrightleftharpoons[k_{2-}]{k_{2+}}$	SB	1
SA	+	B	$\xrightleftharpoons[k_{2-}]{k_{2+}}$	SAB	2
SC	+	B	$\xrightleftharpoons[k_{2-}]{k_{2+}}$	SBC	3
SAC	+	B	$\xrightleftharpoons[k_{2-}]{k_{2+}}$	SABC	4

S	+	C	$\xrightleftharpoons[k_{3-}]{k_{3+}}$	SC	1
SA	+	C	$\xrightleftharpoons[k_{3-}]{k_{3+}}$	SAC	2
SB	+	C	$\xrightleftharpoons[k_{3-}]{k_{3+}}$	SBC	3
SAB	+	C	$\xrightleftharpoons[k_{3-}]{k_{3+}}$	SABC	4

reactions

S, A, B, C,
SA, SB, SC,
SAB, SAC, SBC,
SABC

species

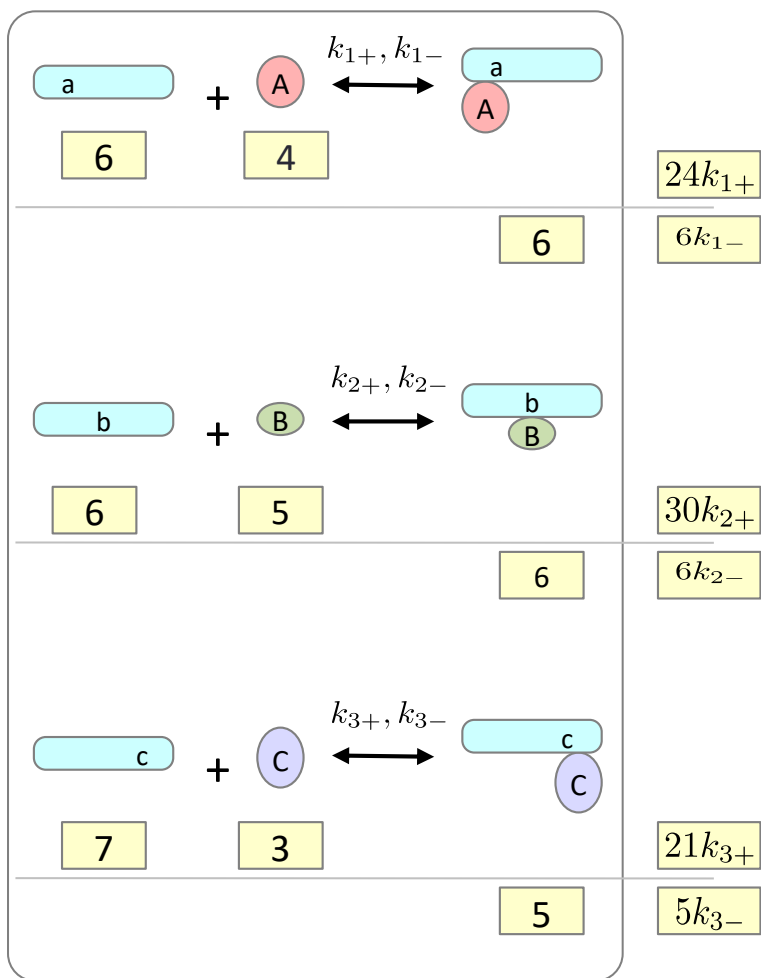
4 molecule types
3 rules



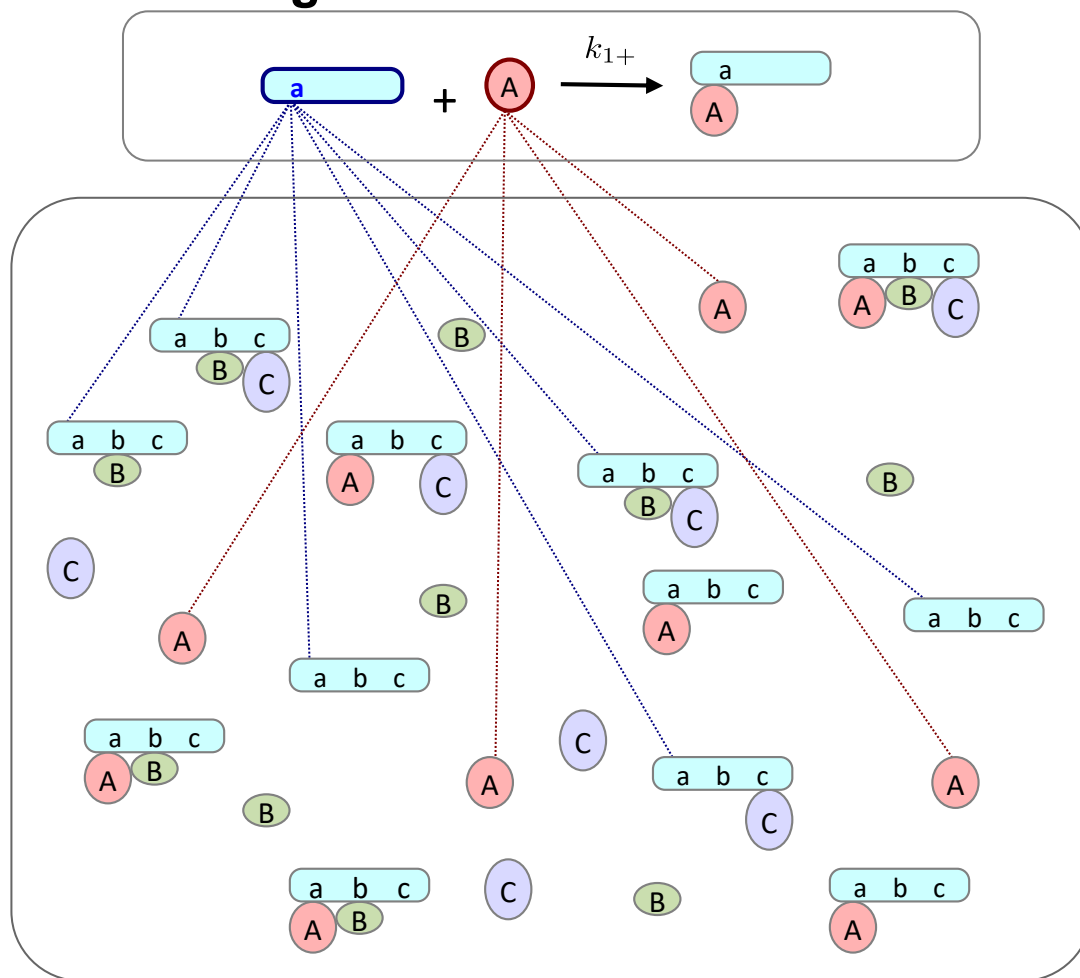
11 species
12 reactions

Direct Methods – rules generate reaction events and system configurations

reaction rules



event generation



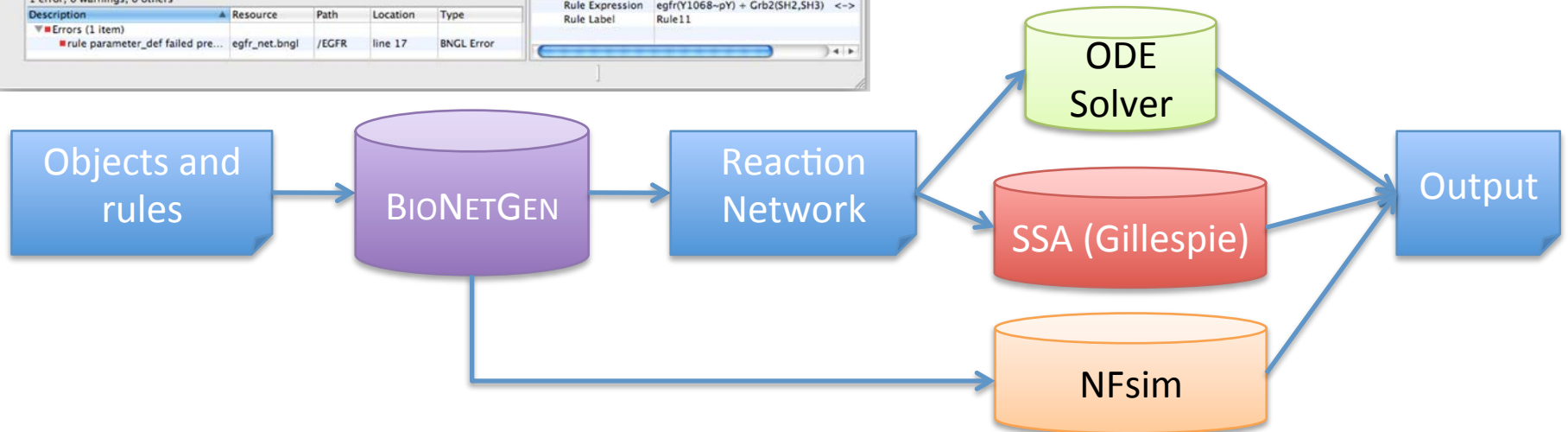
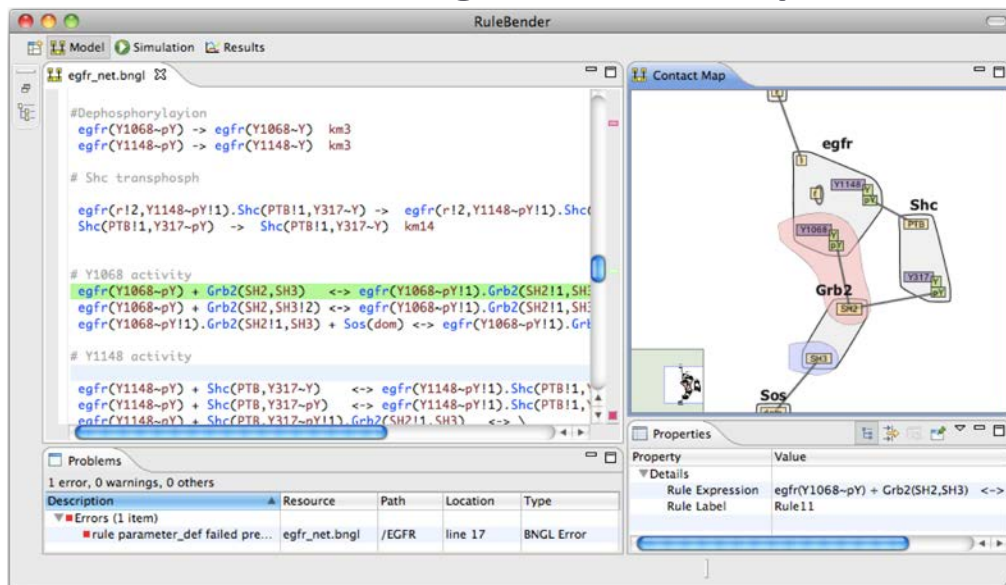
system configuration

total propensity

$$24k_{1+} + 6k_{1-} + 30k_{2+} + 6k_{2-} + 21k_{3+} + 5k_{3-}$$

RuleBender/BioNetGen/NFsim

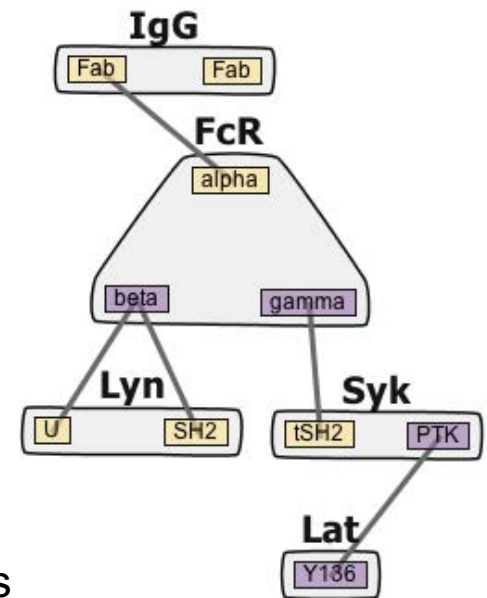
RuleBender – integrated development environment (IDE)



<http://bionetgen.org/index.php/Download>

Why use rule-based modeling techniques?

- **Concise and precise representation of biochemical knowledge**
 - Rules provide a convenient language for representing biomolecular interactions
 - Intricate molecular mechanisms can be captured easily in rule-based models
- **Flexible with respect to simulation method**
 - Deterministic / Stochastic
 - Well-mixed / Compartmental / Spatial
- **Model elements are *modular and reusable***
 - Rule libraries (Chylek et al., 2014) *Frontiers in Immunology*
- **Compact and automatic *visualization***
 - Contact map and beyond
- **Easy *annotation***
 - Model elements can be directly mapped to database entries



Contact map

Outline

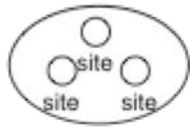
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5. **Exercises (computer lab)**

During the afternoon computer lab (**6/11, Mon**), we will build a simple rule-based model using RuleBender and look at several example models presented in this tutorial/review: Chylek et al. (2015) *Phys Biol* **12**: 045007.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4526164/>

A rule-based model corresponding to the equilibrium continuum model of Goldstein and Perelson (1984)

Molecules



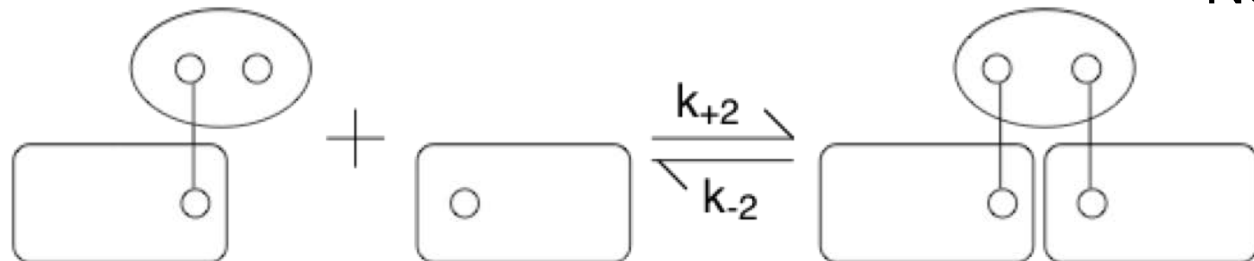
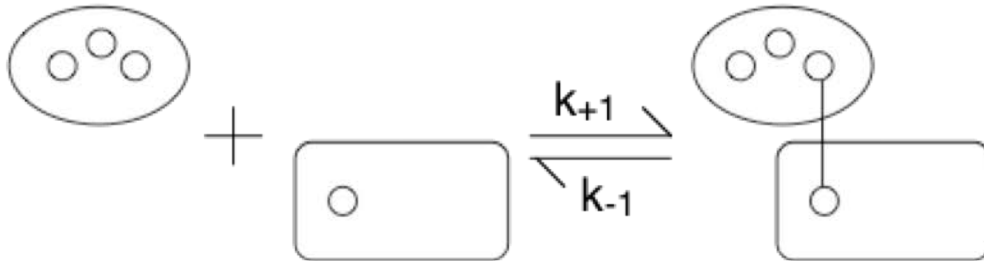
Ligand



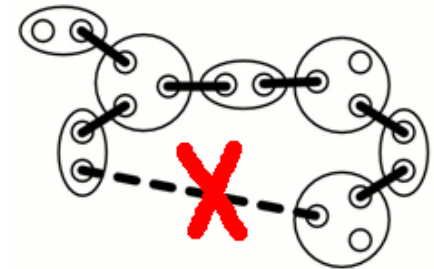
Receptor

This is the “TLBR model.”

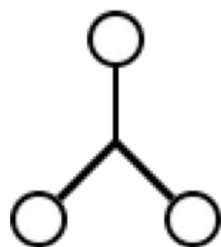
Interactions (reaction rules)



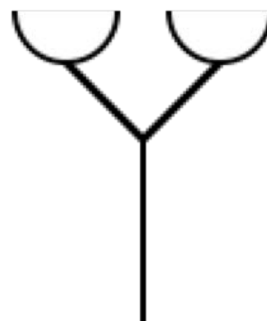
No cyclic aggregates



“Generate-first” method starts with seed species

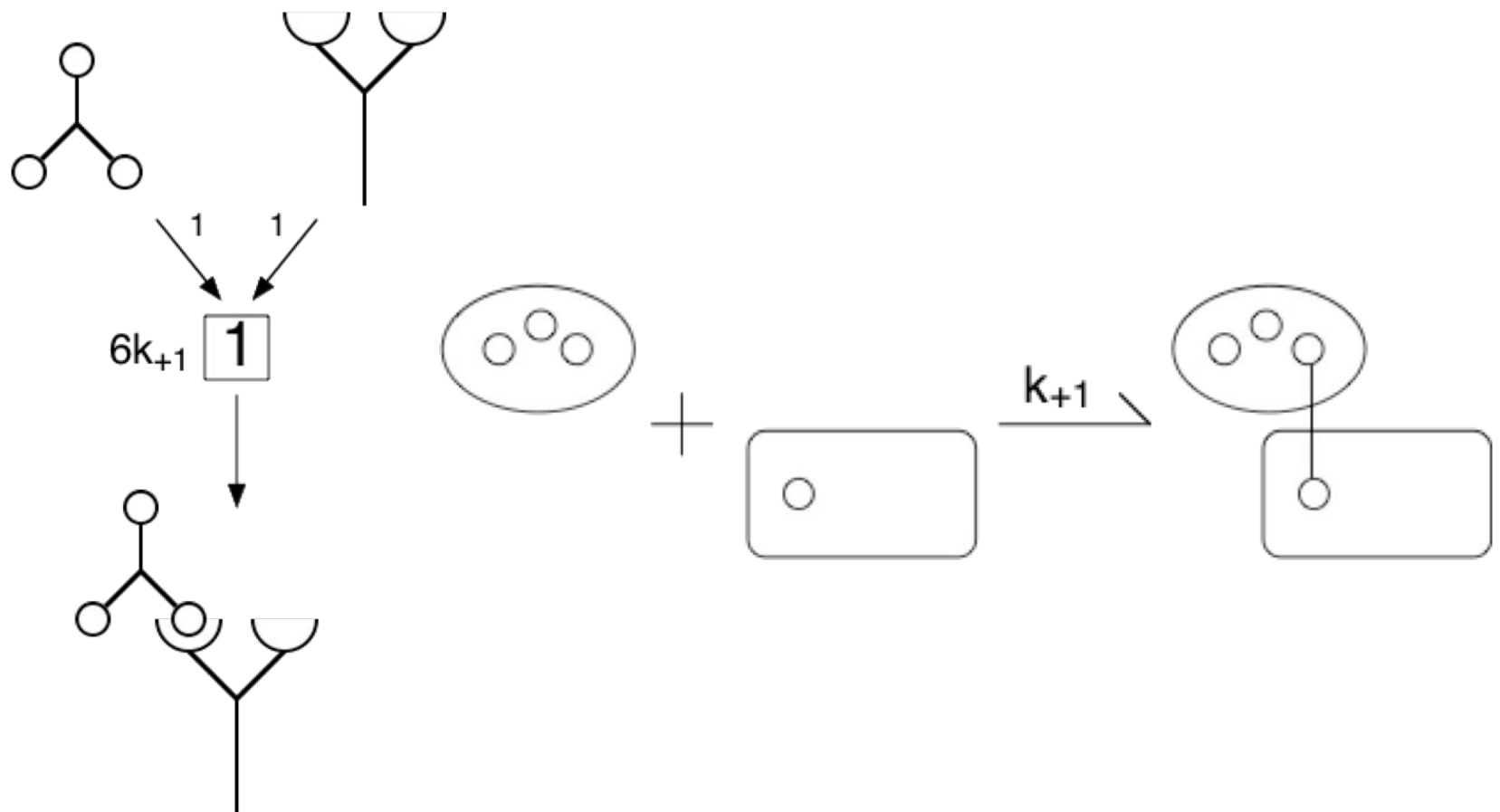


Ligand

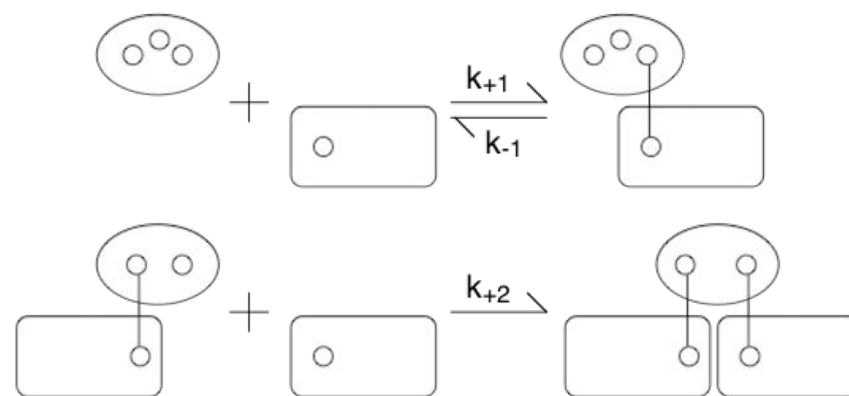
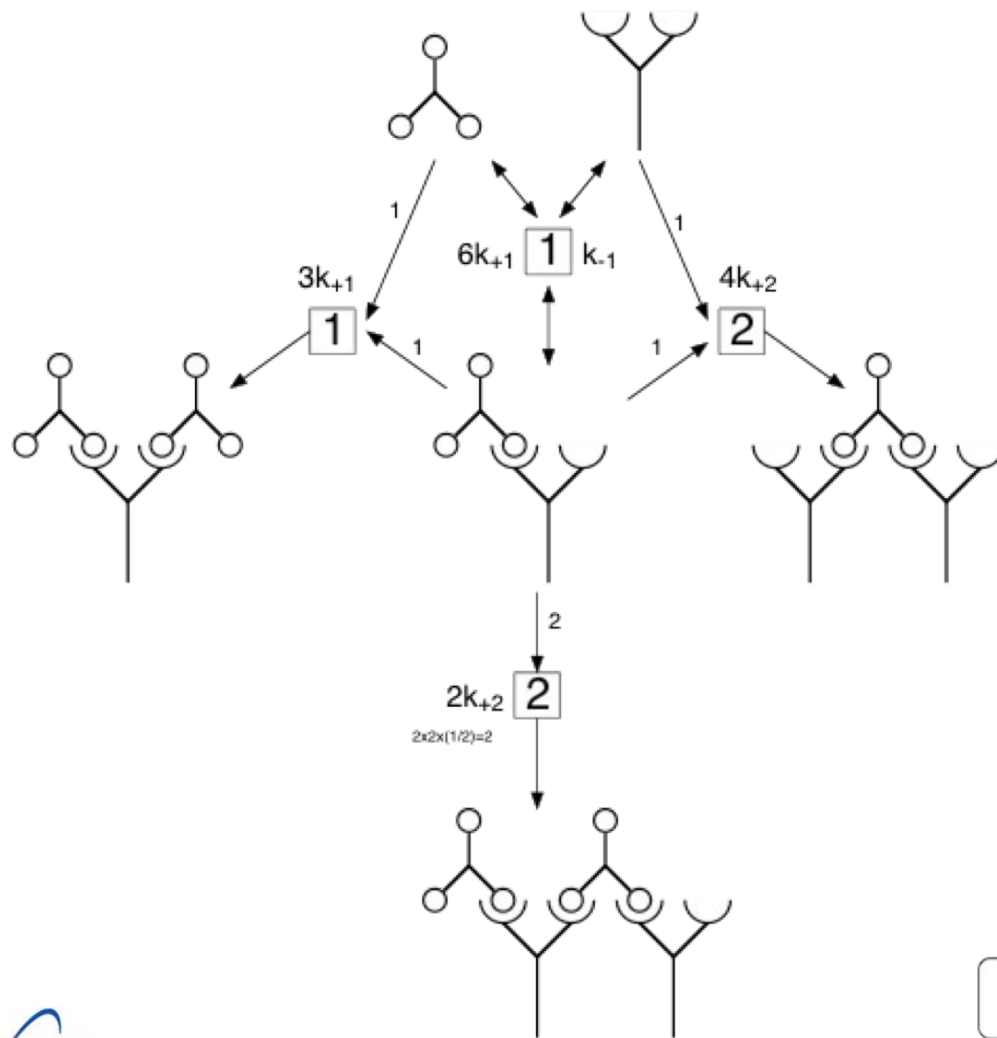


Receptor

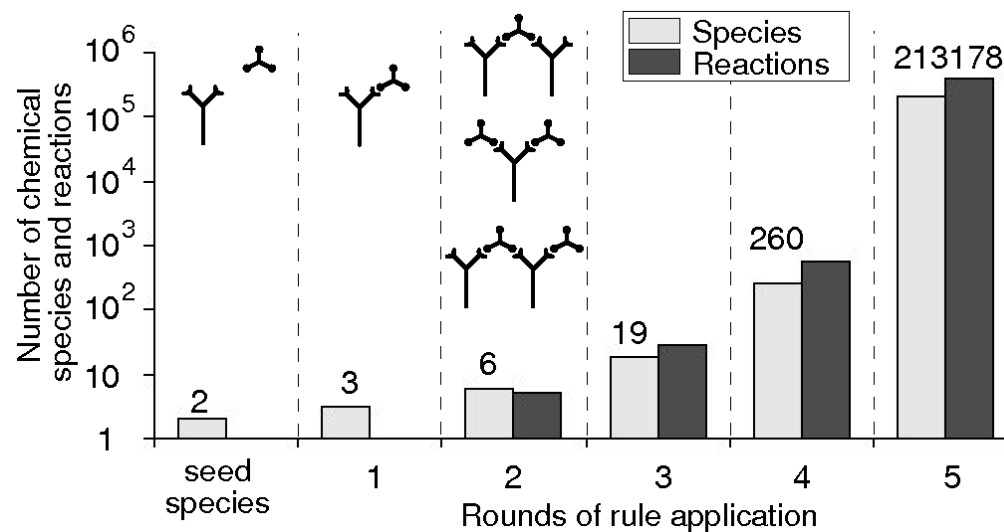
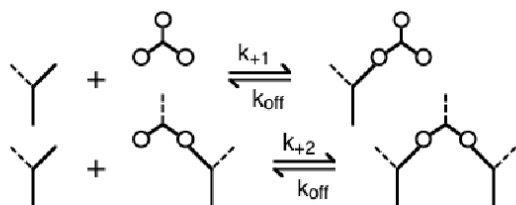
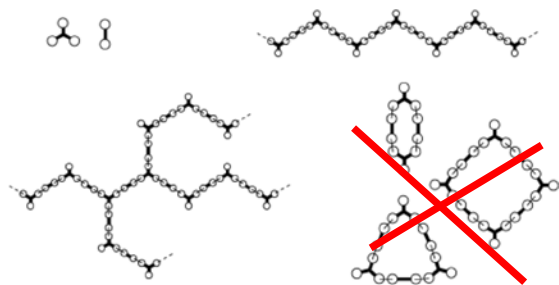
After first round of rule application



After the second round of rule application



Rule-derived network can be too large to simulate using conventional population-based methods



On The Fly method

