

In Search of Lipid Rafts: Evidence of Phospholipid/Cholesterol Complexes

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Interactions at Lipid Interfaces

Human Lung Surfactant

Antimicrobial Peptides

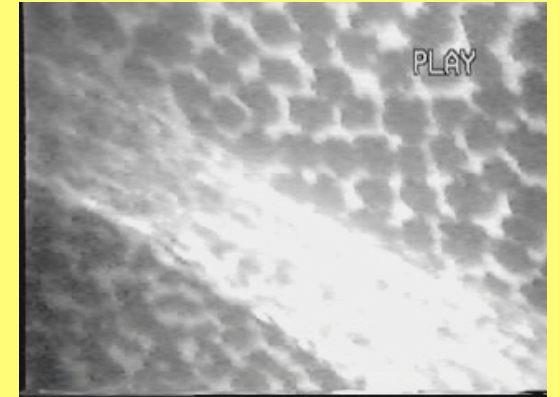
Alzheimer's A β Peptides

Poloxamers as Membrane Sealants

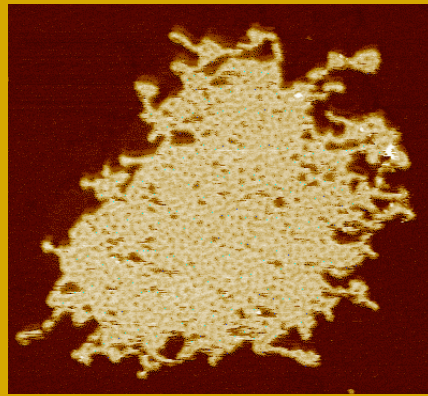
**Phospholipid/Cholesterol
Interactions**

Interactions at Lipid Interfaces

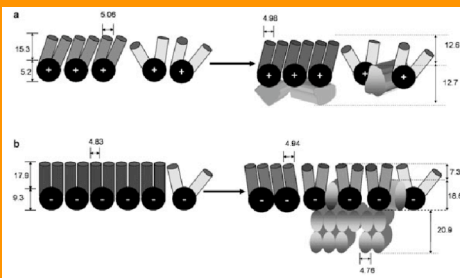
Human Lung Surfactant



Antimicrobial Peptides



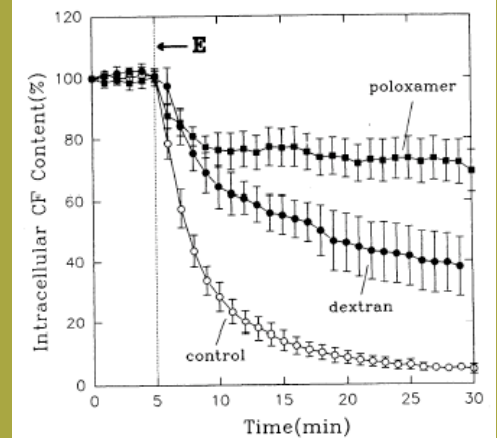
Alzheimer's A β Peptides



Poloxamers as Membrane Sealants

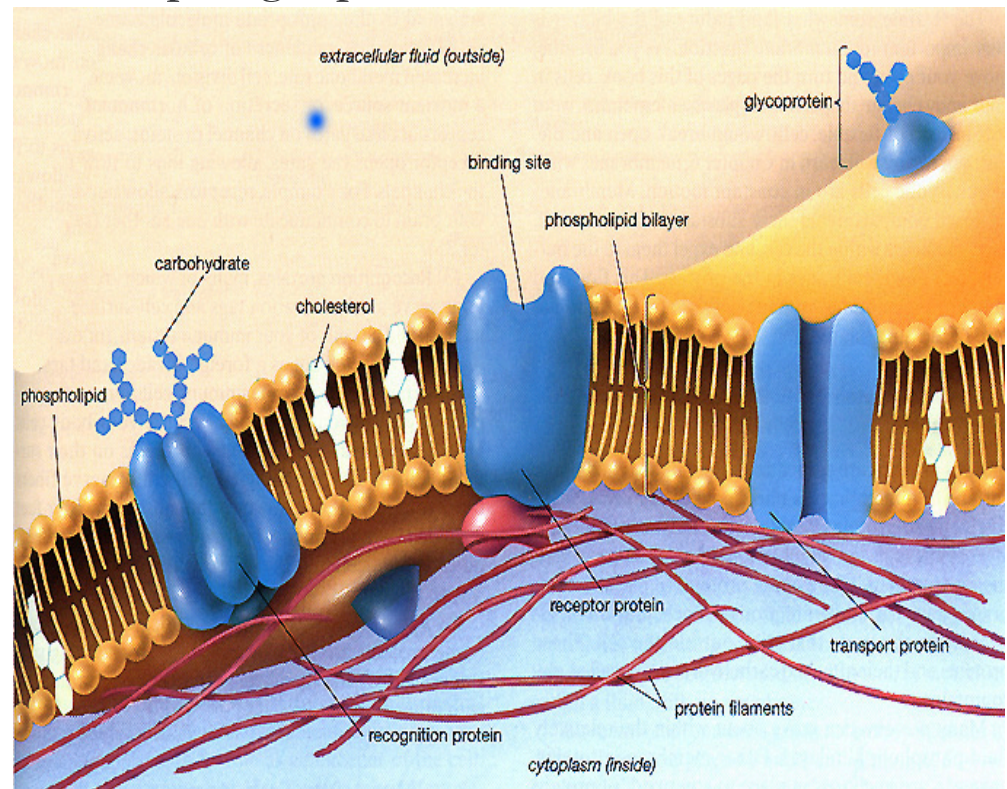


Phospholipid/Cholesterol Interactions



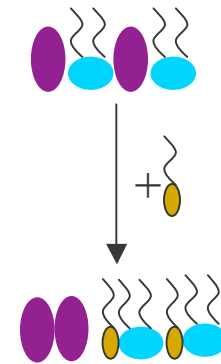
Background

- 1972: Fluid Mosaic Model
- “Lipid raft”: enriched in cholesterol and sphingolipids
Important in signal transduction
- Lengthscale: predicted ~ 100 nm
- Model systems:
GUVs
Monolayers
- Thermodynamic phases?
- Dynamic structures?



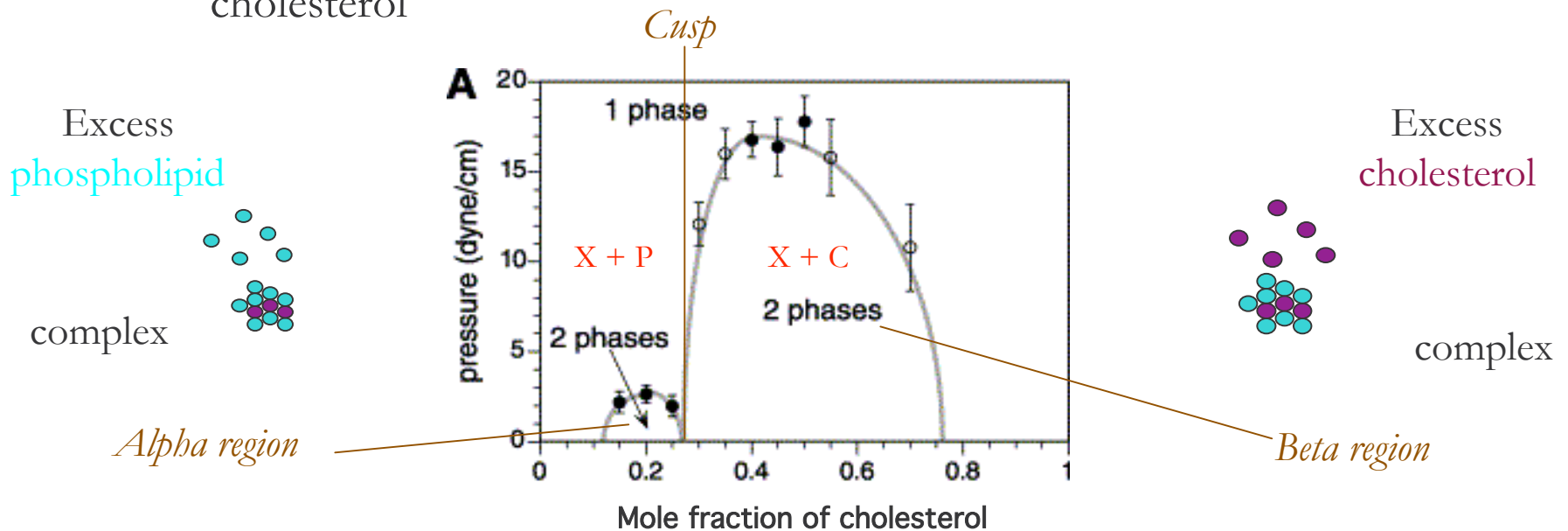
Outline

- Background & Motivation
- Can phospholipid/cholesterol interaction be disrupted?
 - Phase diagrams in the presence of displacing agent
 - Cholesterol desorption by cyclodextrin in monolayers
- Ordering of phospholipid/cholesterol systems
 - Grazing incidence X-ray diffraction and X-ray reflectivity results
- Evidence of displacement in live cell systems
 - "Inhibiting" and "promoting" sterols on cholesterol activity in RBC
- Conclusions



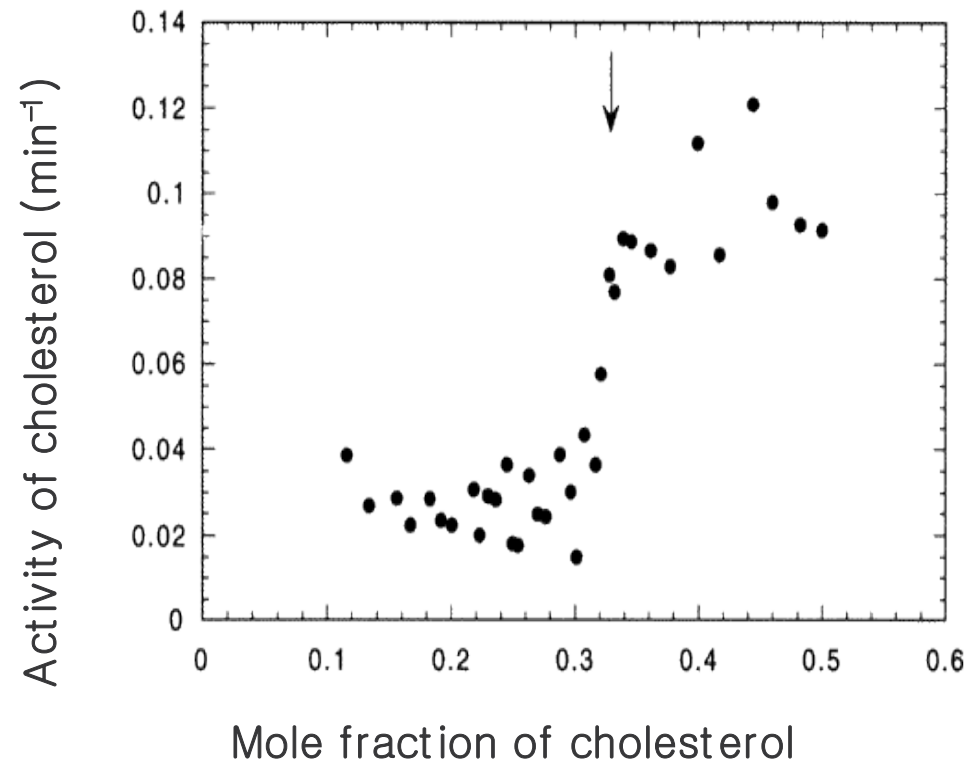
Postulation of Complex Formation

- McConnell postulates the existence of lipid-cholesterol complexes in model membranes
- *Alpha region*: region of low cholesterol concentration; complex + lipid (**X + P**)
- *Beta region*: region of high cholesterol concentration; complex + cholesterol (**X + C**)
- *Cusp*: stoichiometric complex between phospholipid & cholesterol

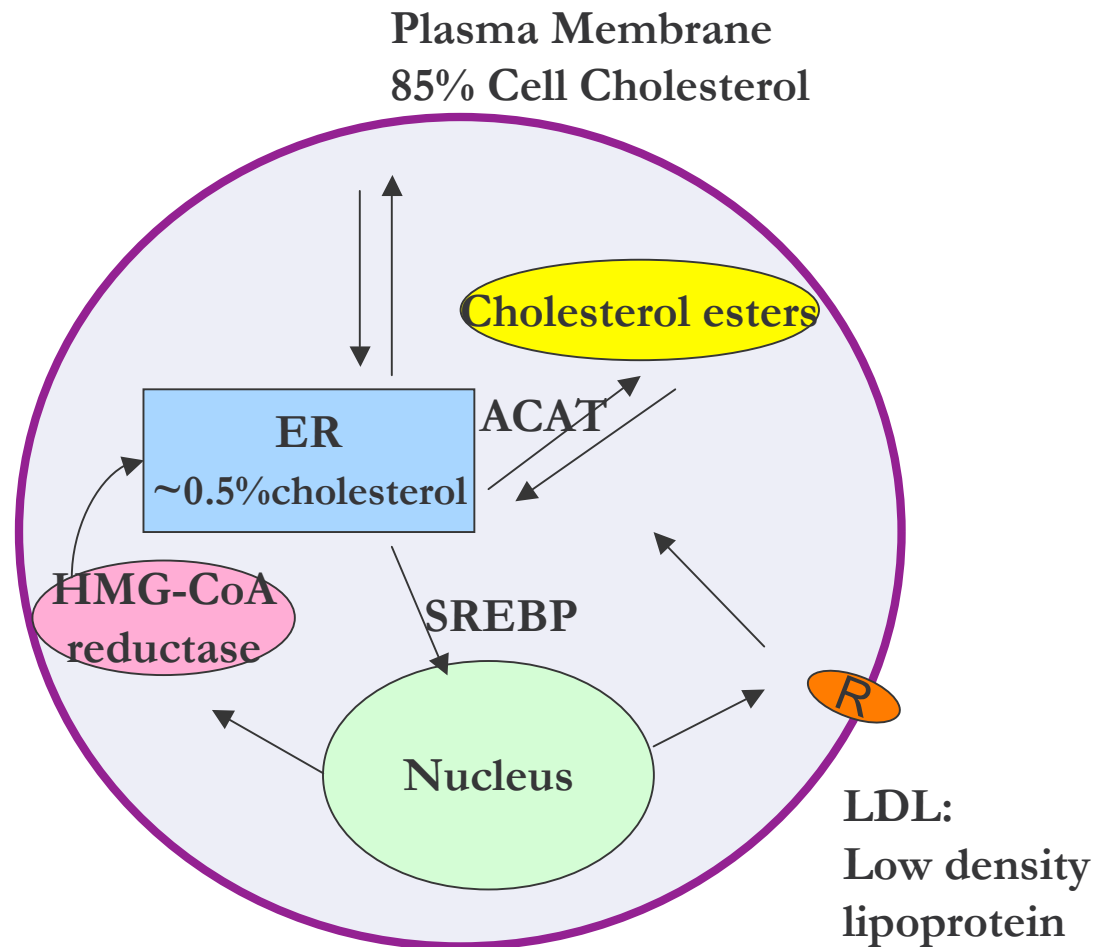


Active (Free) Cholesterol Monomers

Uptake of cholesterol by beta-cyclodextrin (β CD)

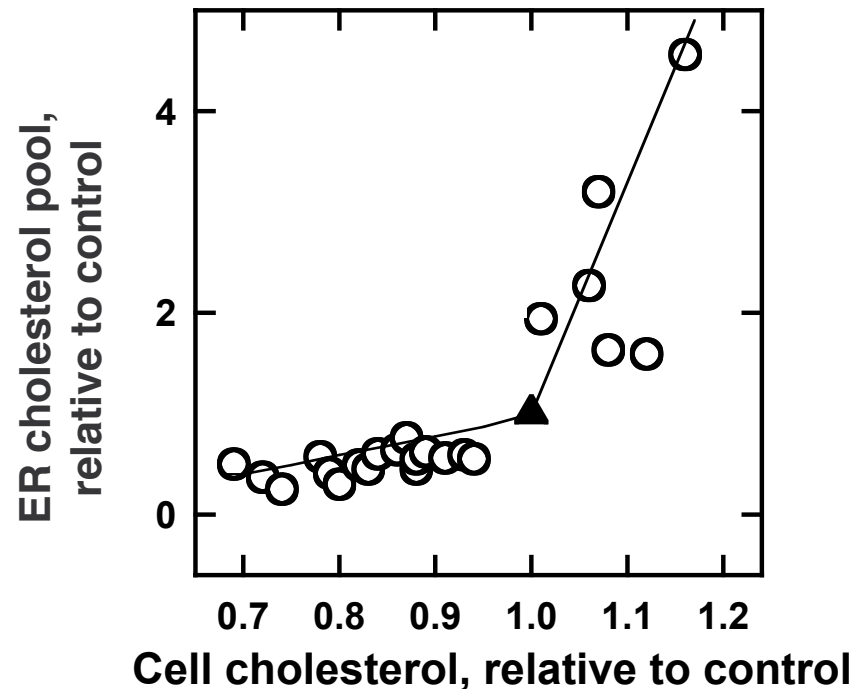


Cholesterol Traffic



Biological Motivation: Cholesterol Homeostasis

- Lange *et al.*: n-octanol can increase cholesterol transfer and ER pool size
- Hypothesis: displacement cholesterol from plasma membrane by n-octanol from the complex, thus freeing up cholesterol monomers
- Understand the J-curve response observed in biological systems

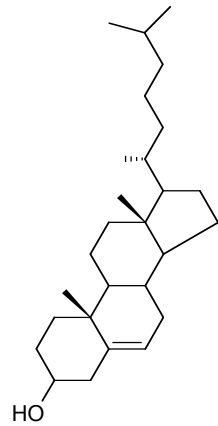


Lange Y, Ye J, Rigney M, Steck TL. (1999) J Lipid Res, 40, 2264-70; also PNAS (2004)

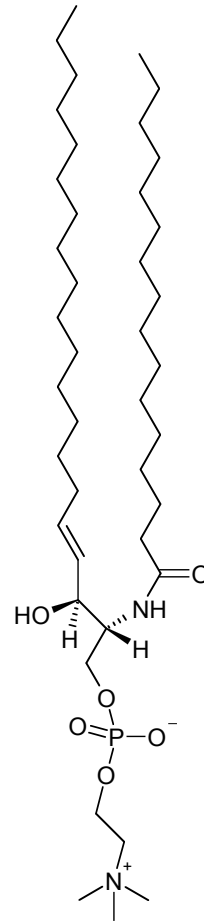
Objectives

- Hypothesis:
 - cholesterol forms **complexes** with phospholipids
- Implications:
 - Cholesterol can exist in two different states:
 - bound cholesterol** (*low activity*)
 - free cholesterol** (*high activity*)
 - Other molecules that can form tighter complexes with phospholipids can be **displacing agents**, freeing cholesterol from its bound state
- Mechanistic Model
 - **cholesterol activity modulation by specific displacing agents.**

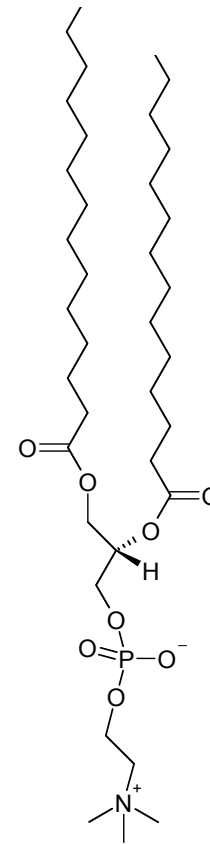
Lipid Systems



Cholesterol



Typical SM



DMPC

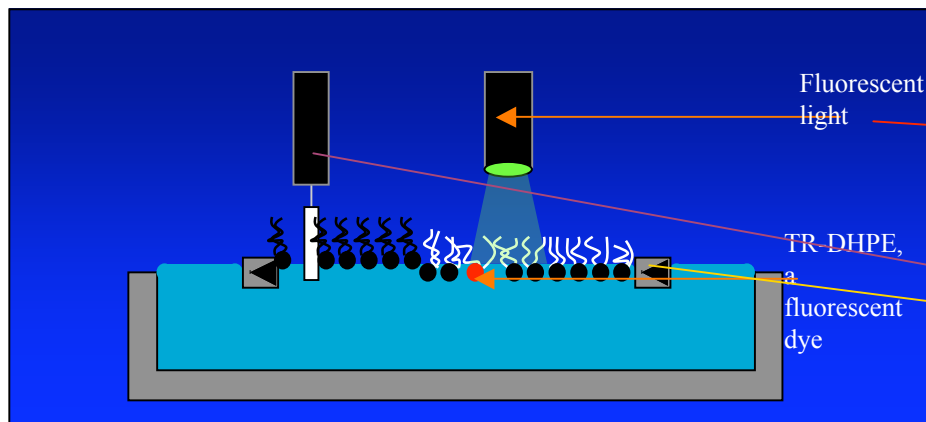
Four Different Approaches

- Phase diagrams
- Cholesterol desorption by beta-cyclodextrin (CD)
- X-ray diffraction and x-ray reflectivity
- Effect of "inhibiting" and "promoting" sterols on cholesterol activity in RBC

Four Different Approaches

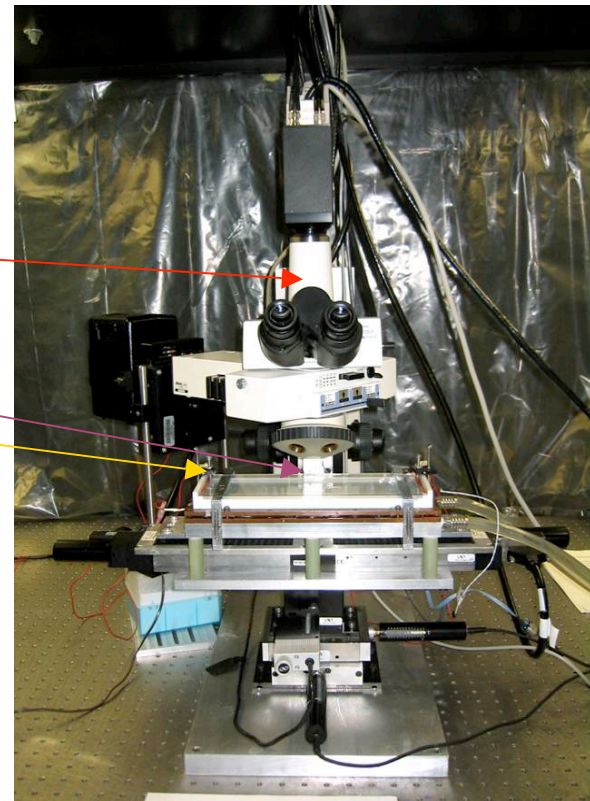
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Experimental Techniques



$$\Pi = \gamma_0 - \gamma$$

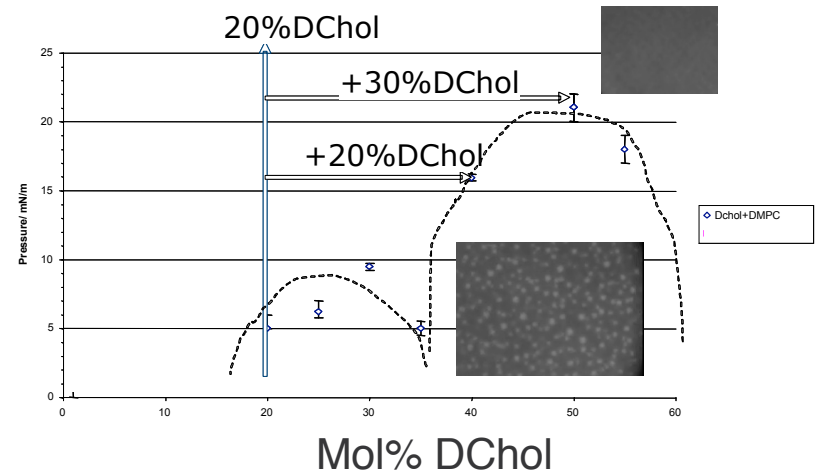
A = Area per molecule



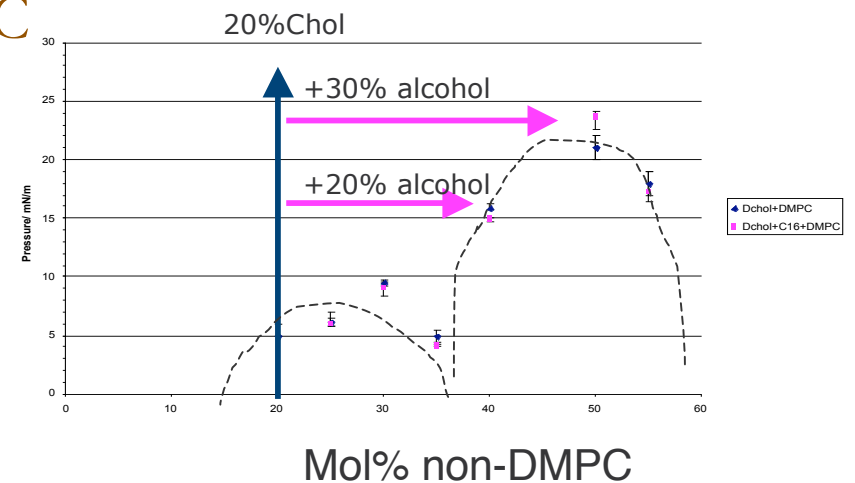
Fluorescence Langmuir Surface Balance

Phase Diagram: Point Shifting

1. DChol:DMPC



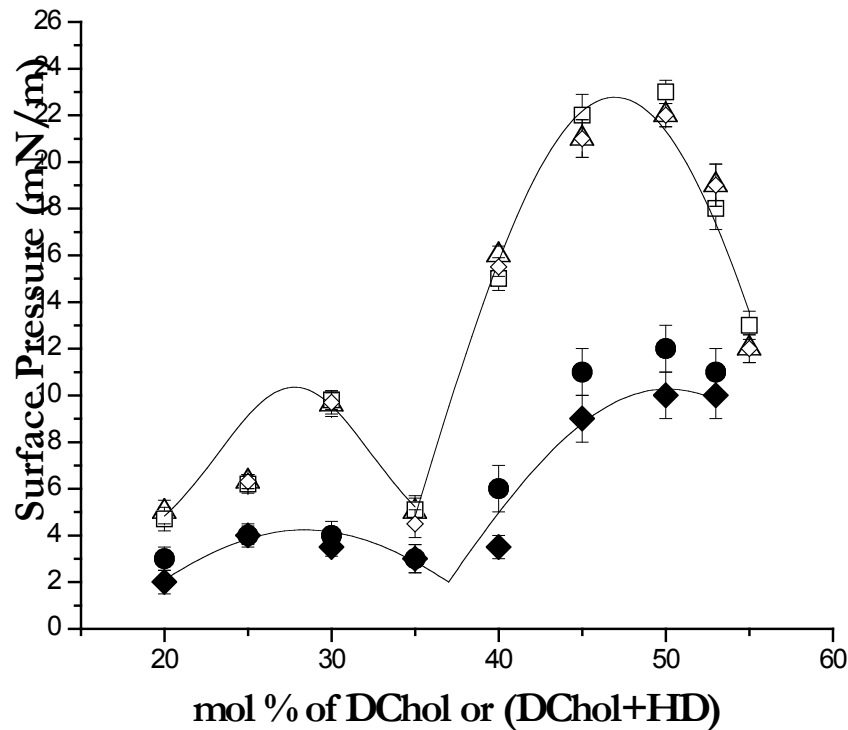
2. 20%DChol:Hexadecanol (C16):DMPC



3. C16:DMPC

No liquid-liquid immiscibility observed

Cholesterol fixed at different mole fractions



Open square 0% DChol
Filled circle 10% DChol
Filled diamond 15% DChol
Open triangle 20% DChol
Open circle 25% DChol

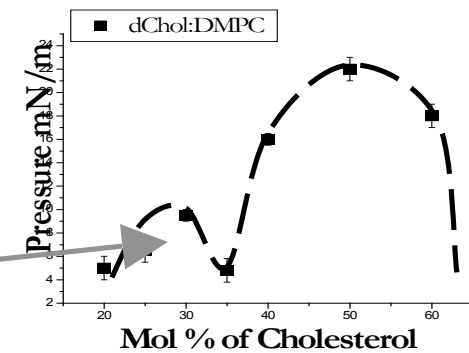
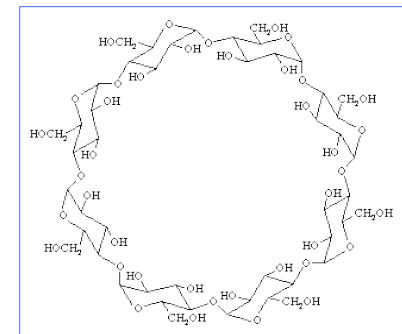
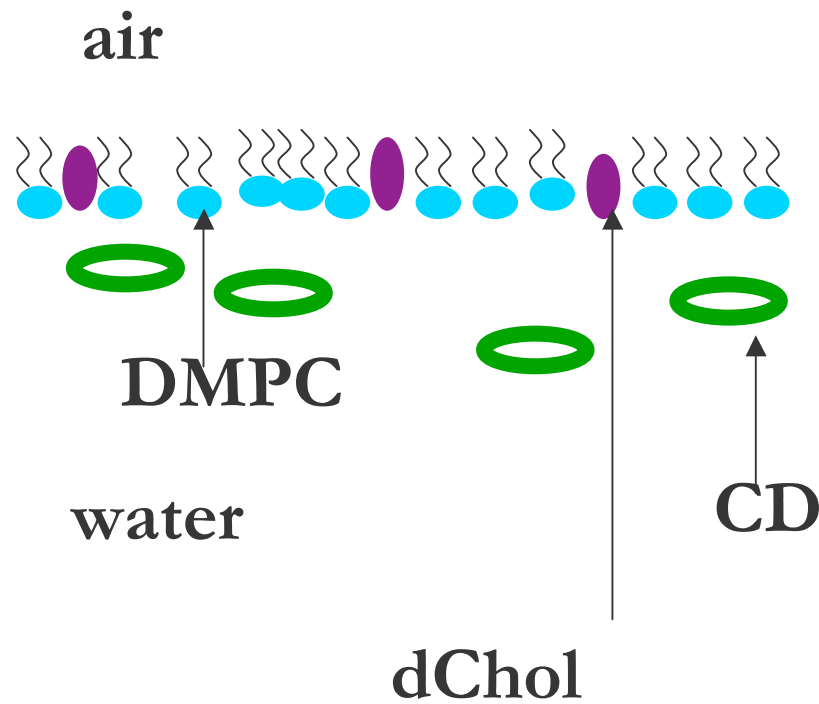
Conclusions from Phase Diagram Study

- Phase diagrams in the presence of displacing agent is equivalent to ones with just pure cholesterol, even at low cholesterol content
- Displacement of cholesterol from its association with phospholipid by alcohol
- The displacement seems to be 1 C16:1 DChol, since we observe identical phase diagrams
- **Does displacement results in the release of free cholesterol?**

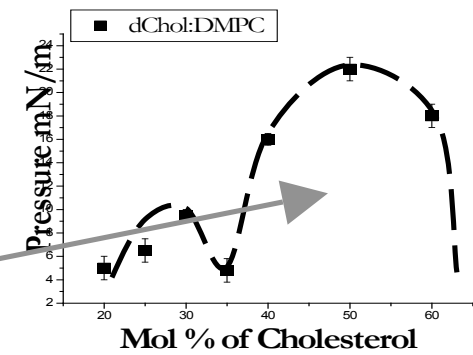
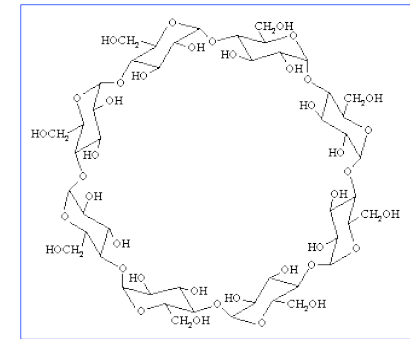
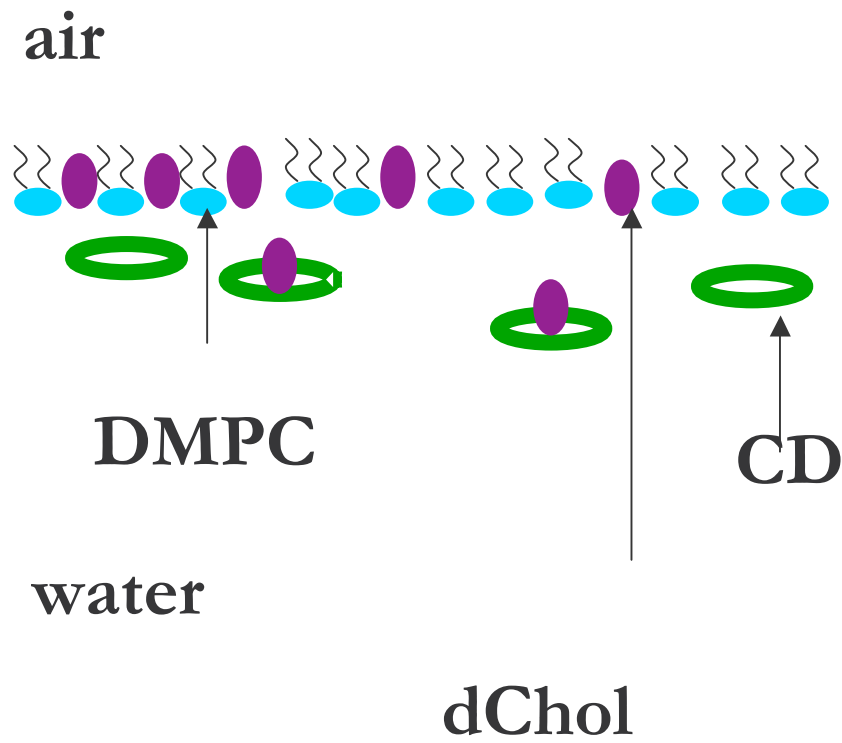
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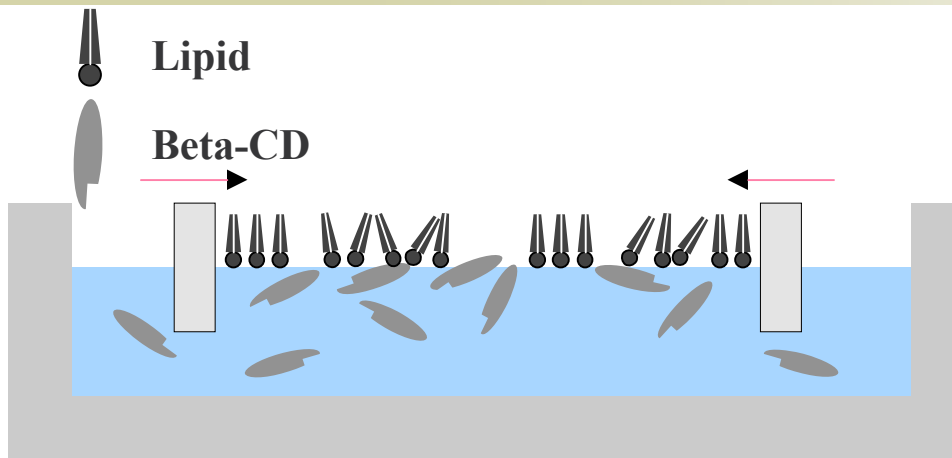
Cholesterol Transfer to CD



Cholesterol Transfer to CD



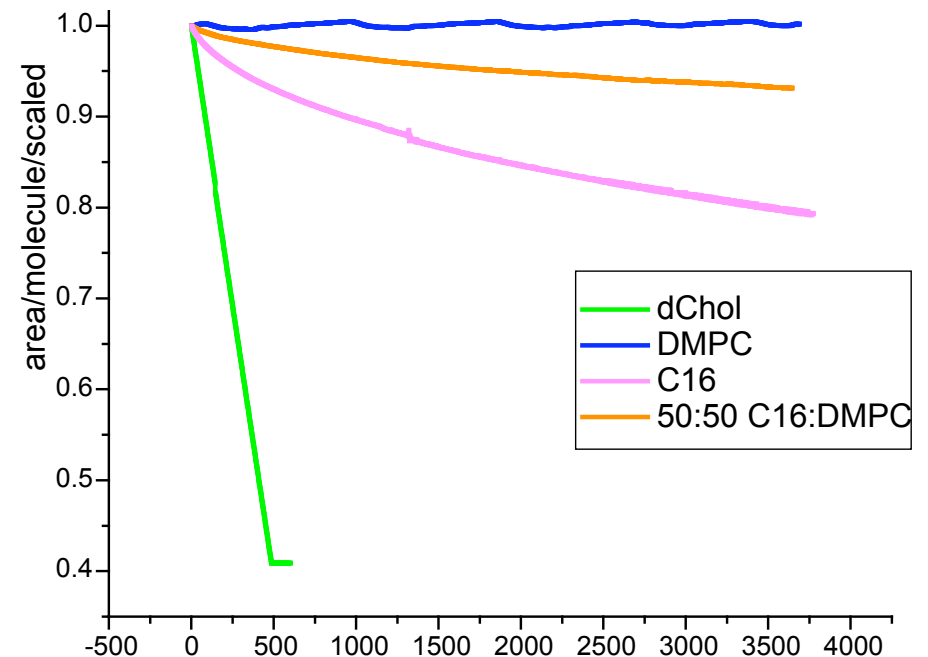
Selectivity of Cyclodextrin



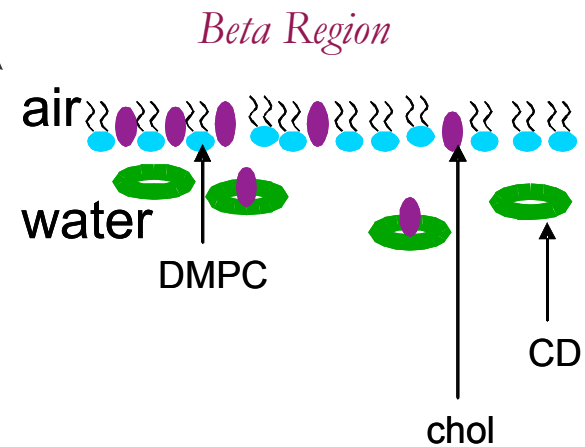
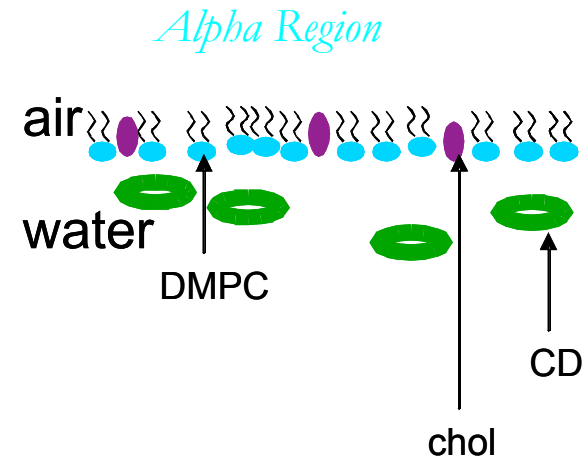
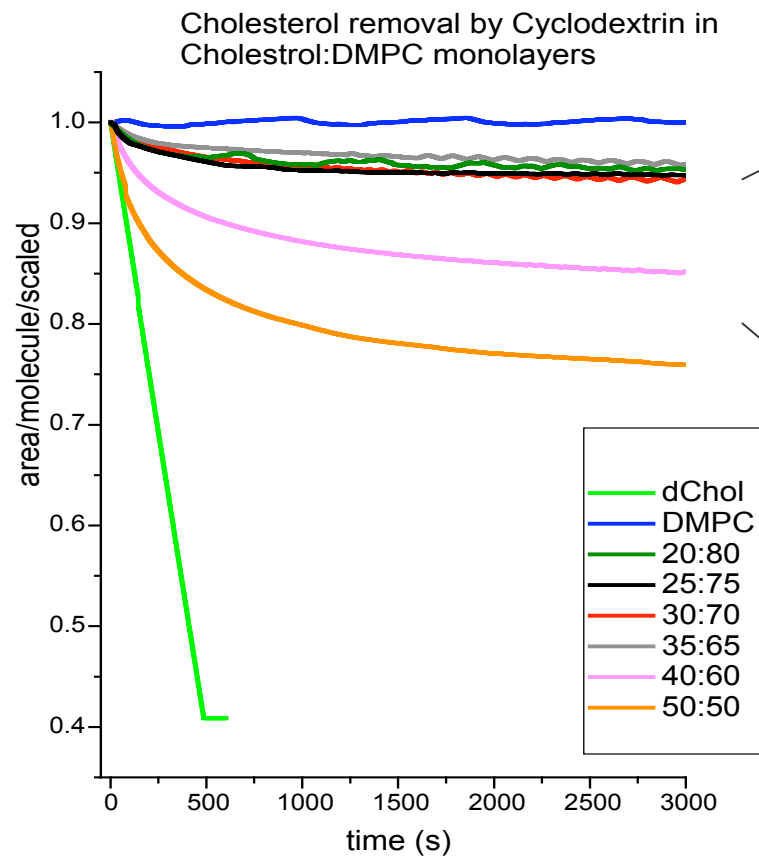
- β -CD removes sterols from monolayers
- Desorption rate of **cholesterol** to CD is much greater than the rates for **DMPC** or **hexadecanol (C16)**

1. Monolayer Compression
memb. equiv. $\Pi \approx 28$ mN/m
(20~35 mN/m)
2. Beta-CD in subphase
3. Area Contraction

Cholesterol removal by Cyclodextrin in
Cholesterol:DMPC monolayers

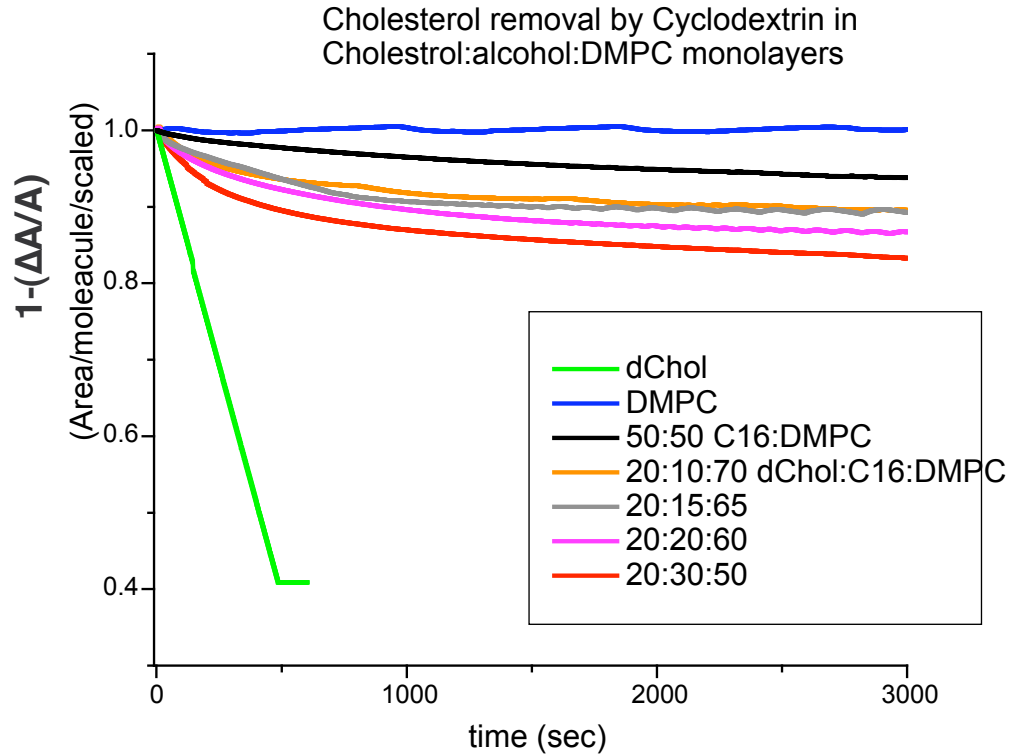


Cyclodextrin on System 1: DChol:DMPC



Cyclodextrin on System 2

20% DChol: C16:DMPC



- Ternary mixtures displays similar behavior to β CD as binary mixtures of DChol:DMPC
- Even with equally low cholesterol content we see changes in desorption rates indicating that alcohol is displacing cholesterol
- Compare **black** and **red** curves

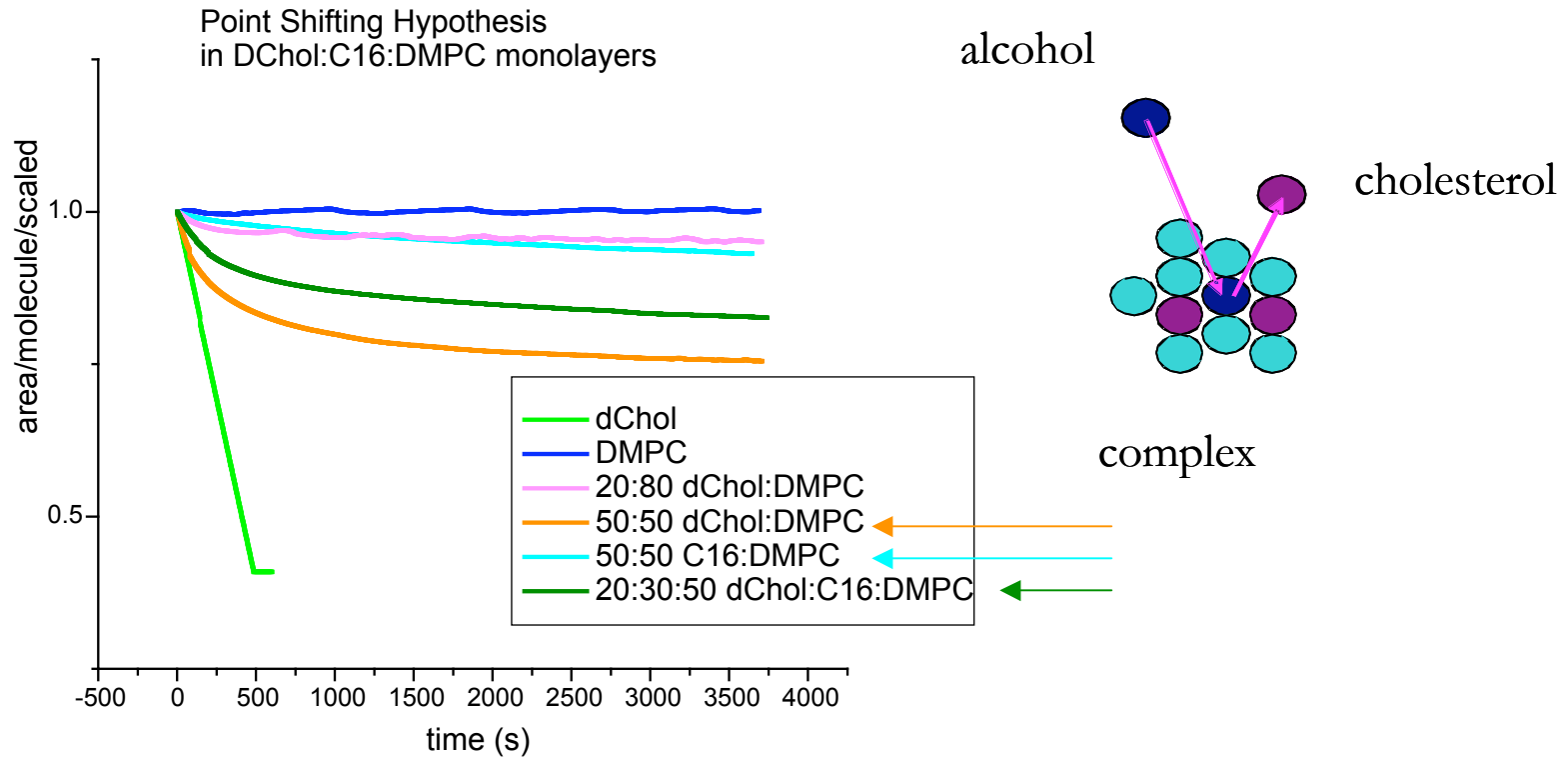
50:50

C16:DMPC

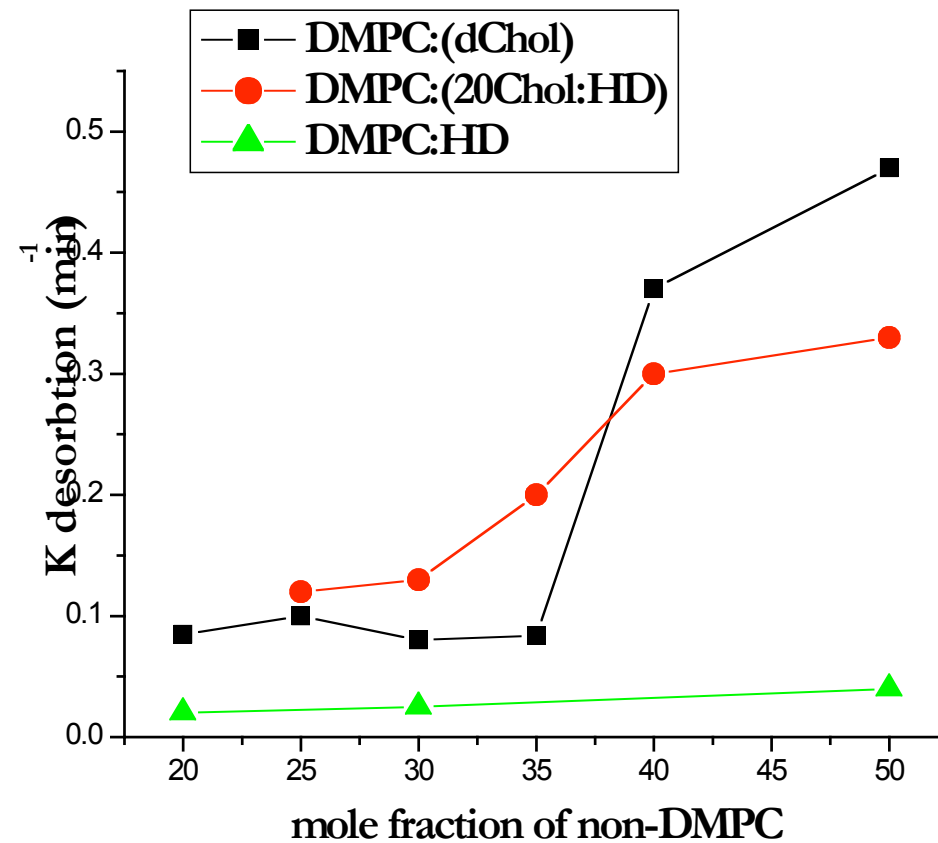
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DChol:C16:DMPC

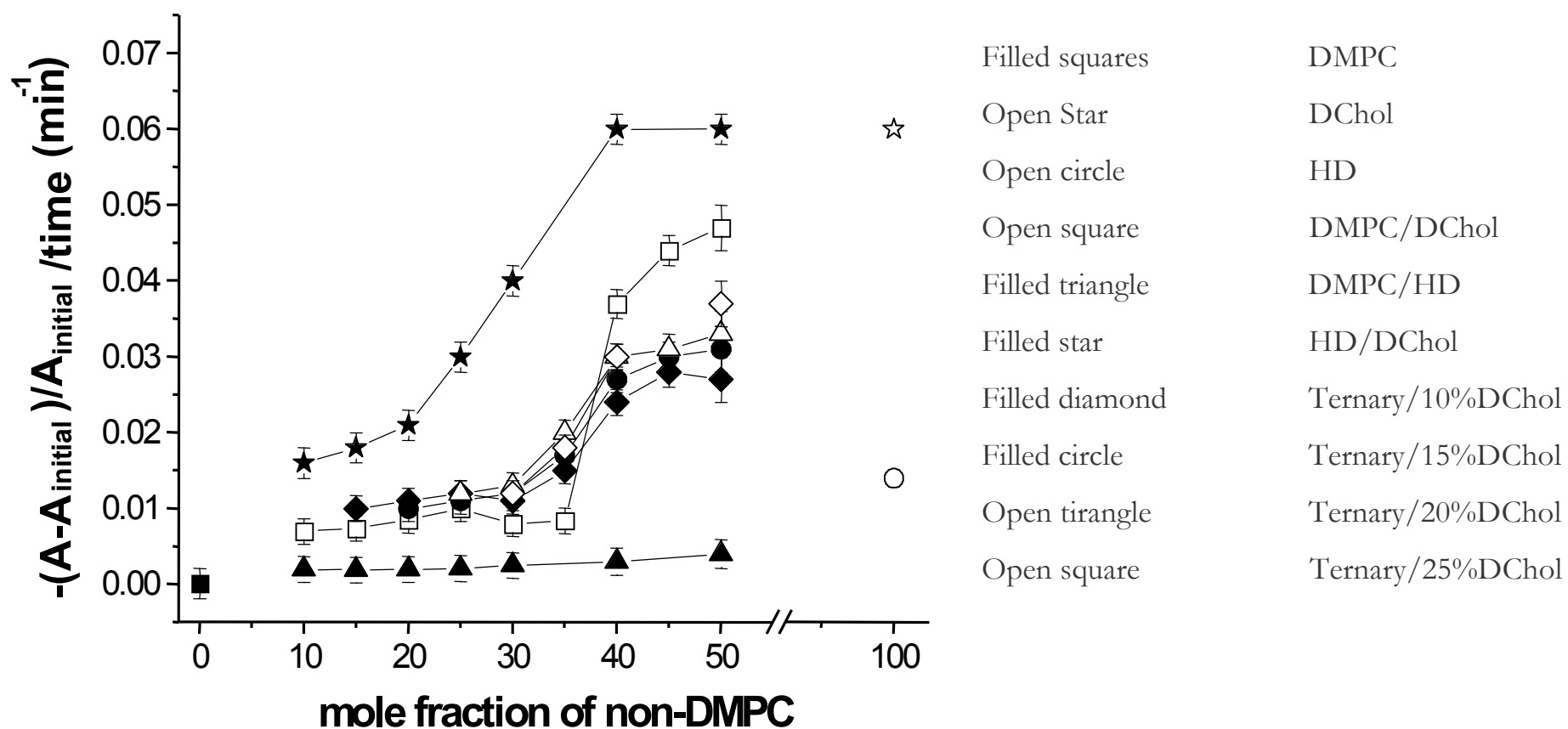
Cyclodextrin II: point shifting



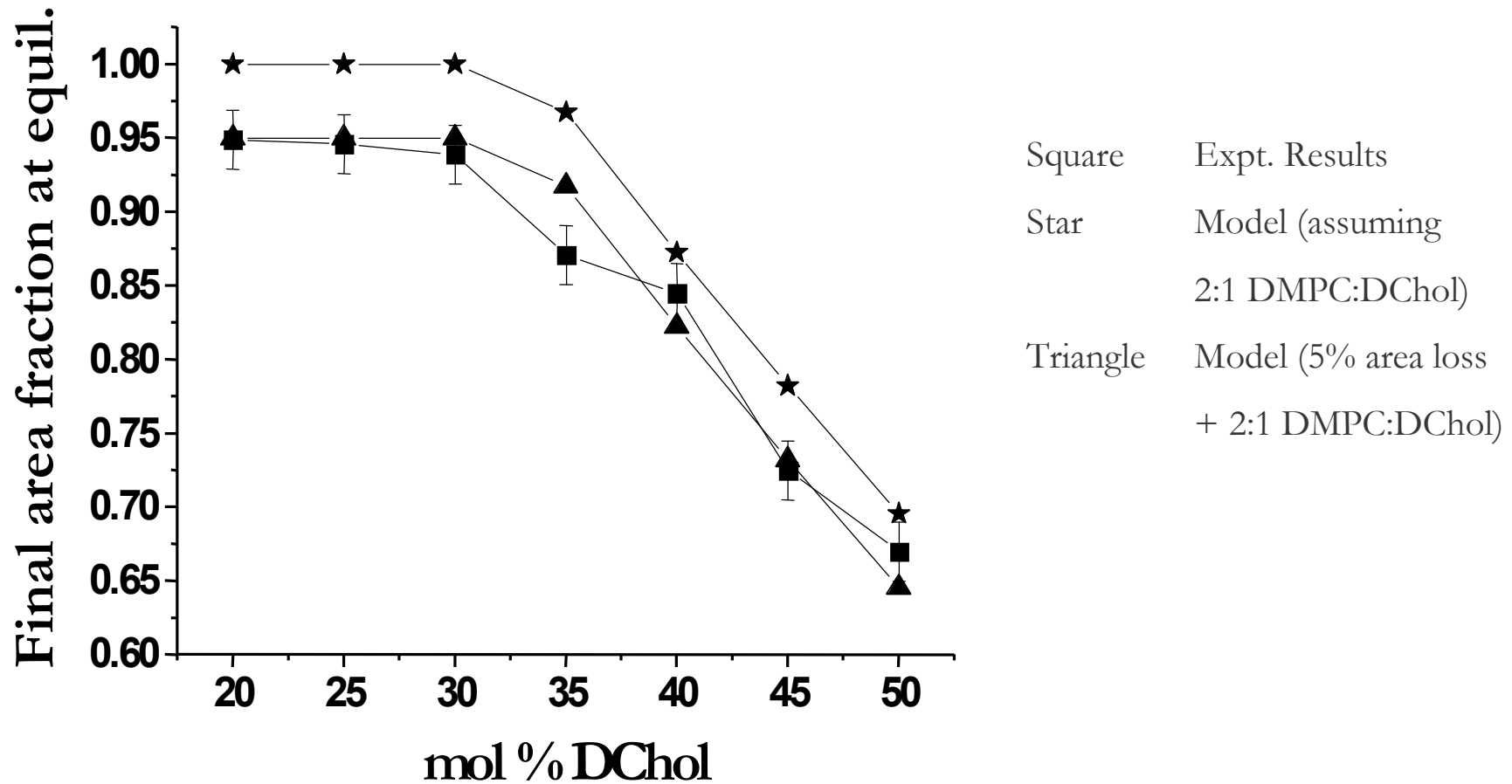
Cholesterol Transfer to CD



Cholesterol Fixed at Different Mole Fractions



Final Equilibrium Area for DMPC/DChol Mixtures



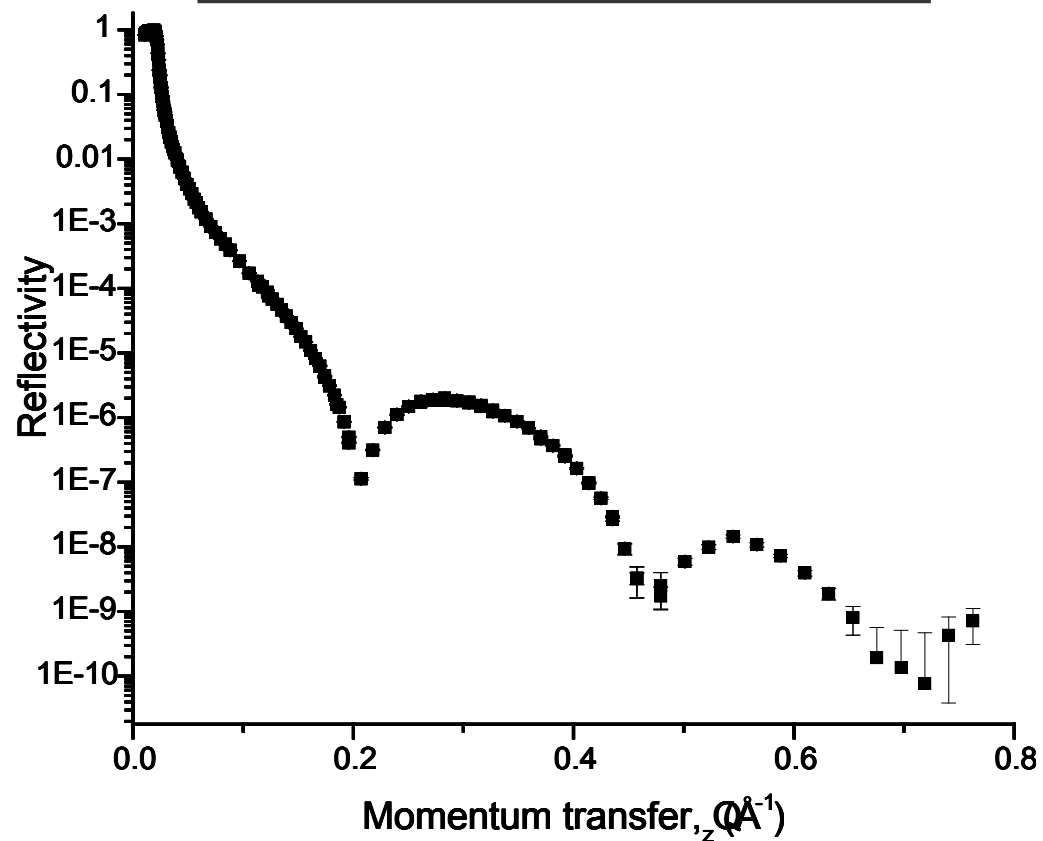
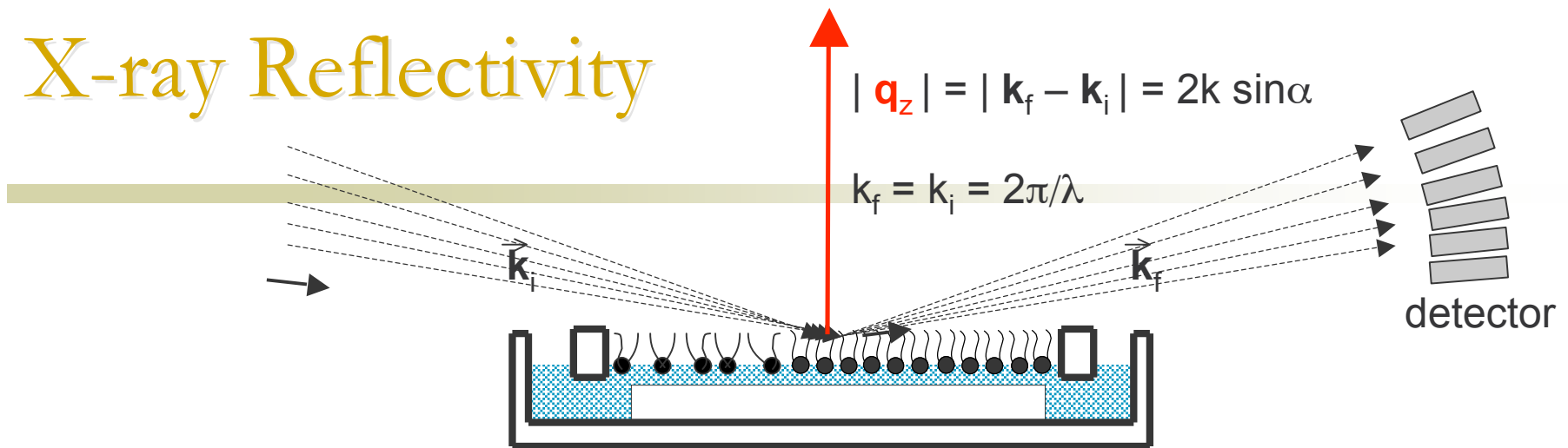
Conclusions from CD Desorption Study

- Cholesterol can exist in two states of low and high chemical potential/activity/fugacity
- **In the presence of displacing agent cholesterol activity is increased**

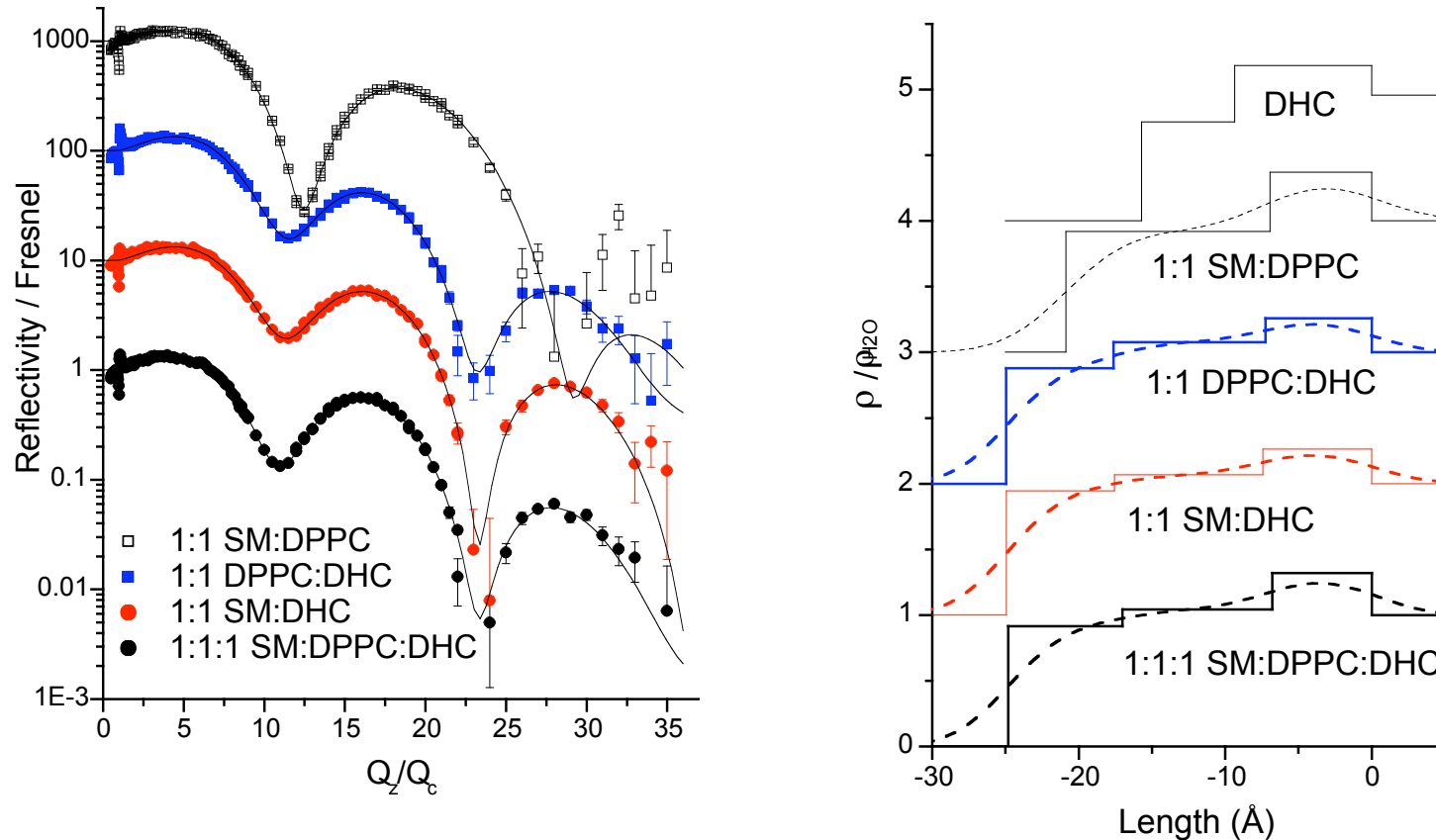
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X-ray Reflectivity

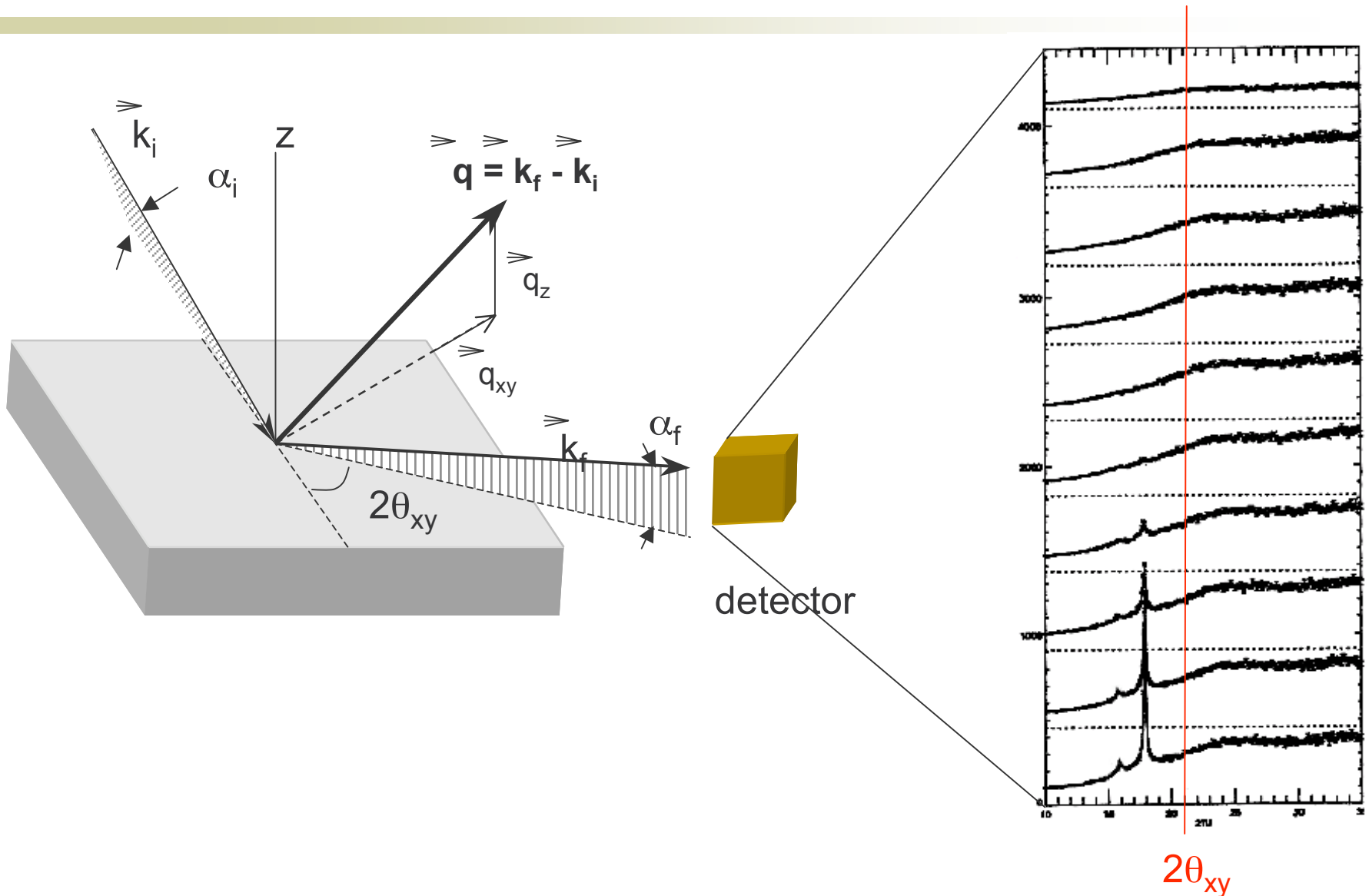


XR Data on Mixtures

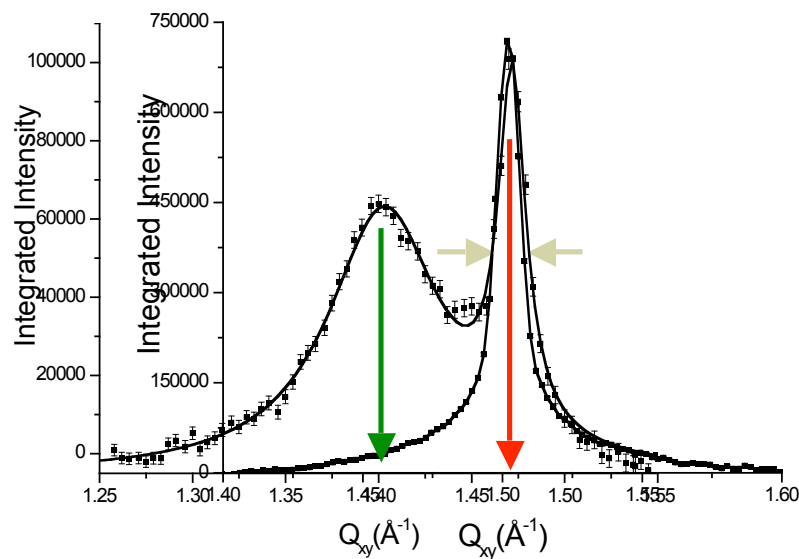
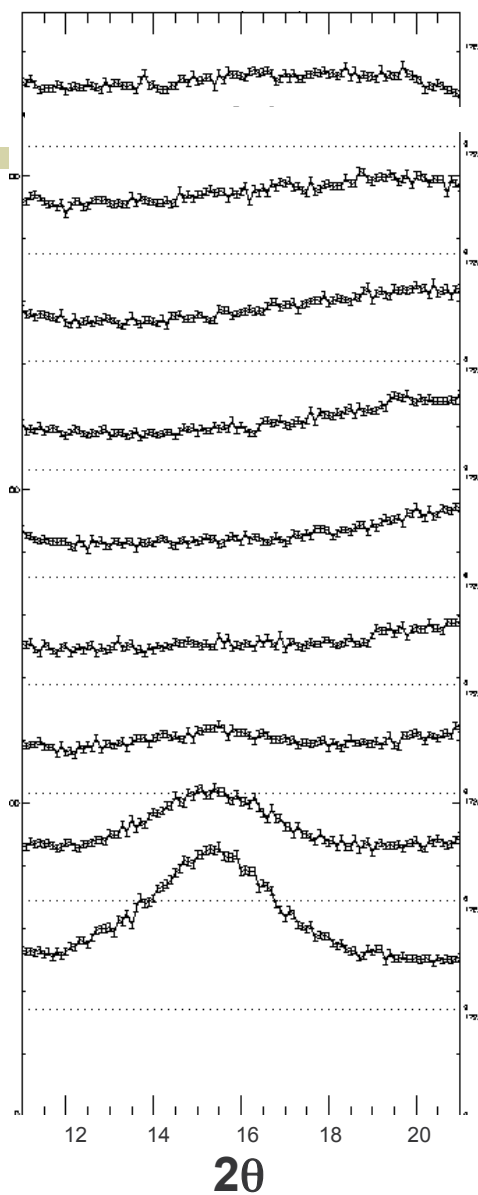


- DChol intercalates into acyl chain region (10 Å) - this layer is also responsible for Bragg scattering observed in GIXD)
- Reduces the tilt of phospholipids (cholesterol condensation effects)

Grazing Incidence X-ray Diffraction (GIXD)



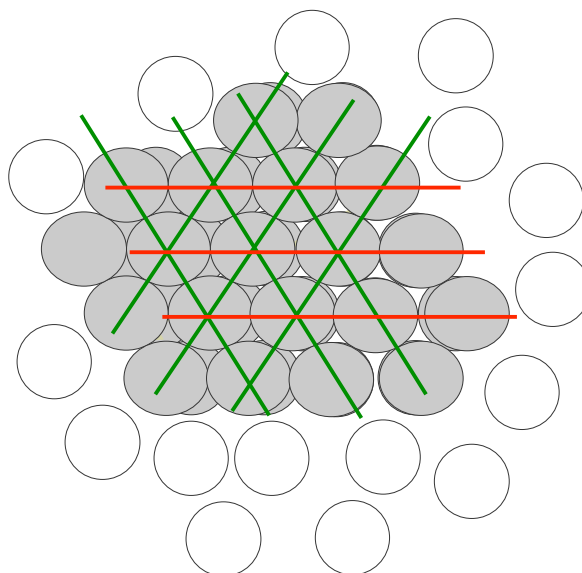
band averaged intensity



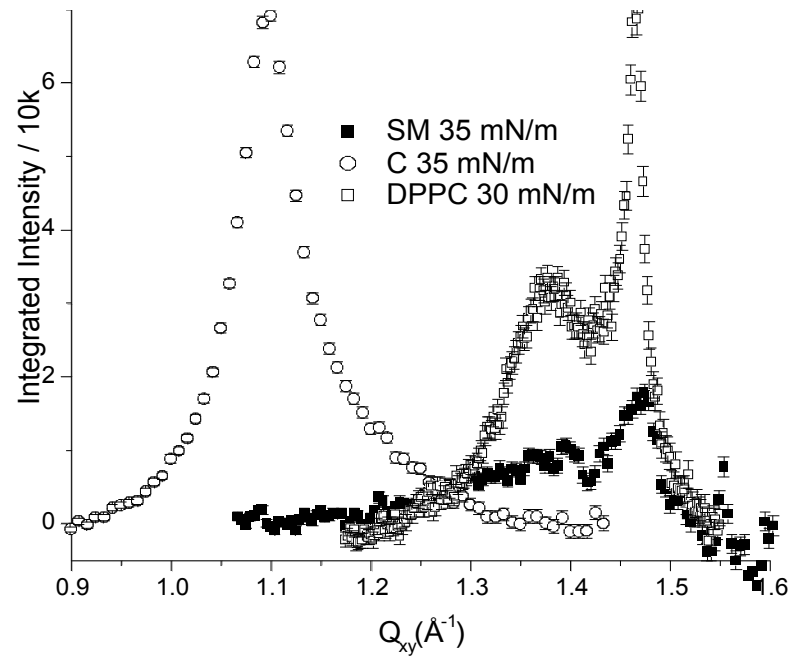
GIXD

$q_{xy} \sim 1 / \text{d-spacing}$

$\text{FWHM} \sim 1 / \text{coh. length}$

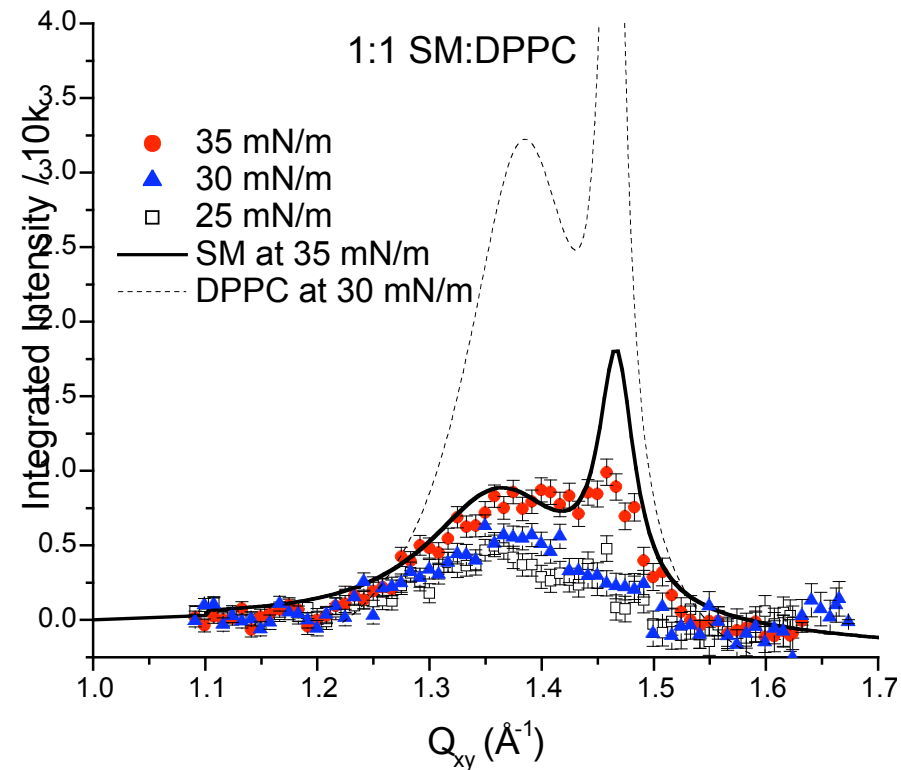


GIXD of Pure Components



	d-spacing(s) (\AA)	Coherence Lengths (\AA) L11, L10 01	Integrated Intensity
SM (35 mN/m)	4.29, 4.61	173,40	0.5
DChol (35 mN/m)	5.72	74.6	1.5
DPPC (30 mN/m)	4.29, 4.56	242, 47	1.3

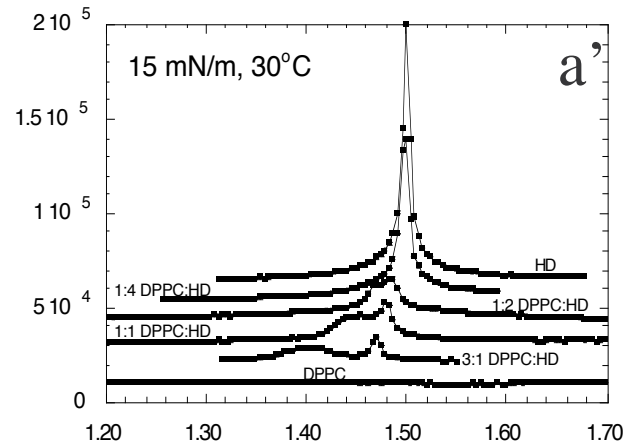
Pressure Dependence of Phospholipid Systems



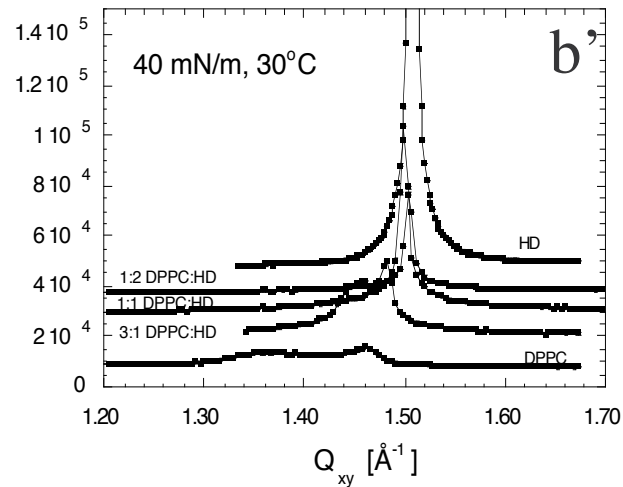
d-spacing, scattering intensity, coherence length (memory of crystallinity) are pressure-dependent

Effects of C16 on Lipid Structure

Low pressure



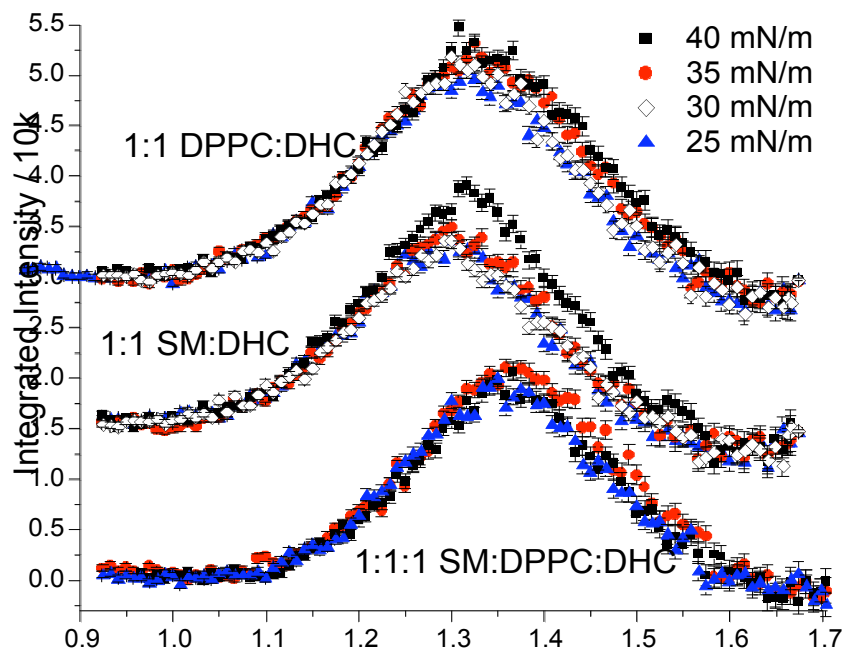
High pressure



KYCL et al.
J. Chem. Phys.
116 (2002) 774-783

- d-spacing, scattering intensity, coherence length (memory of crystallinity) are pressure-dependent
- C16 induces lipid packing at a lower pressure

Presence of DChol Leads to Very Different Scattering Behavior



- coherence length $\sim 22 \text{ \AA}$
- d-spacing $\neq f$ (surface pressure)
- coherence length $\neq f$ (surface pressure)
- amt of scattering entities $\neq f$ (surface pressure)
- DChol = DHC

1:1 DPPC:DChol

Π	d spacing	coherence length	integrated intensity
25	4.79	22.37	1.1
30	4.79	22.61	1.2
35	4.75	21.46	1.2
40	4.73	21.01	1.4

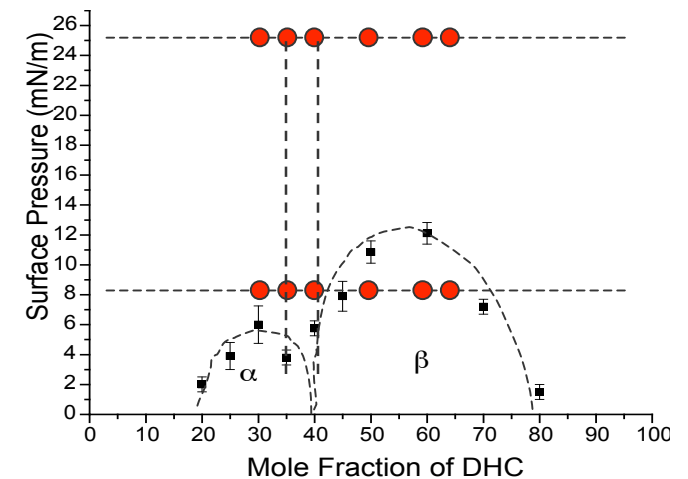
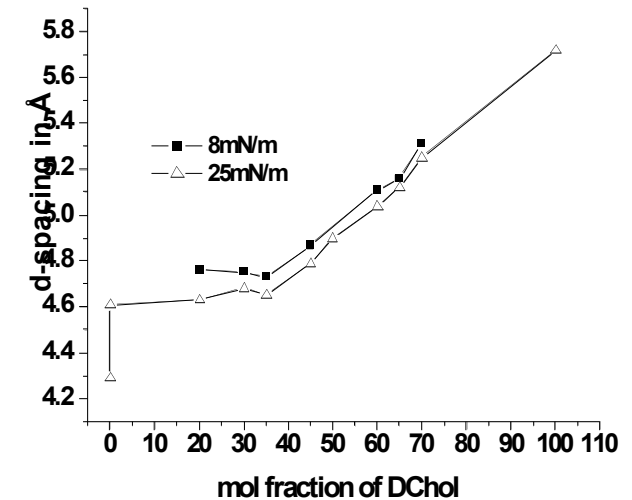
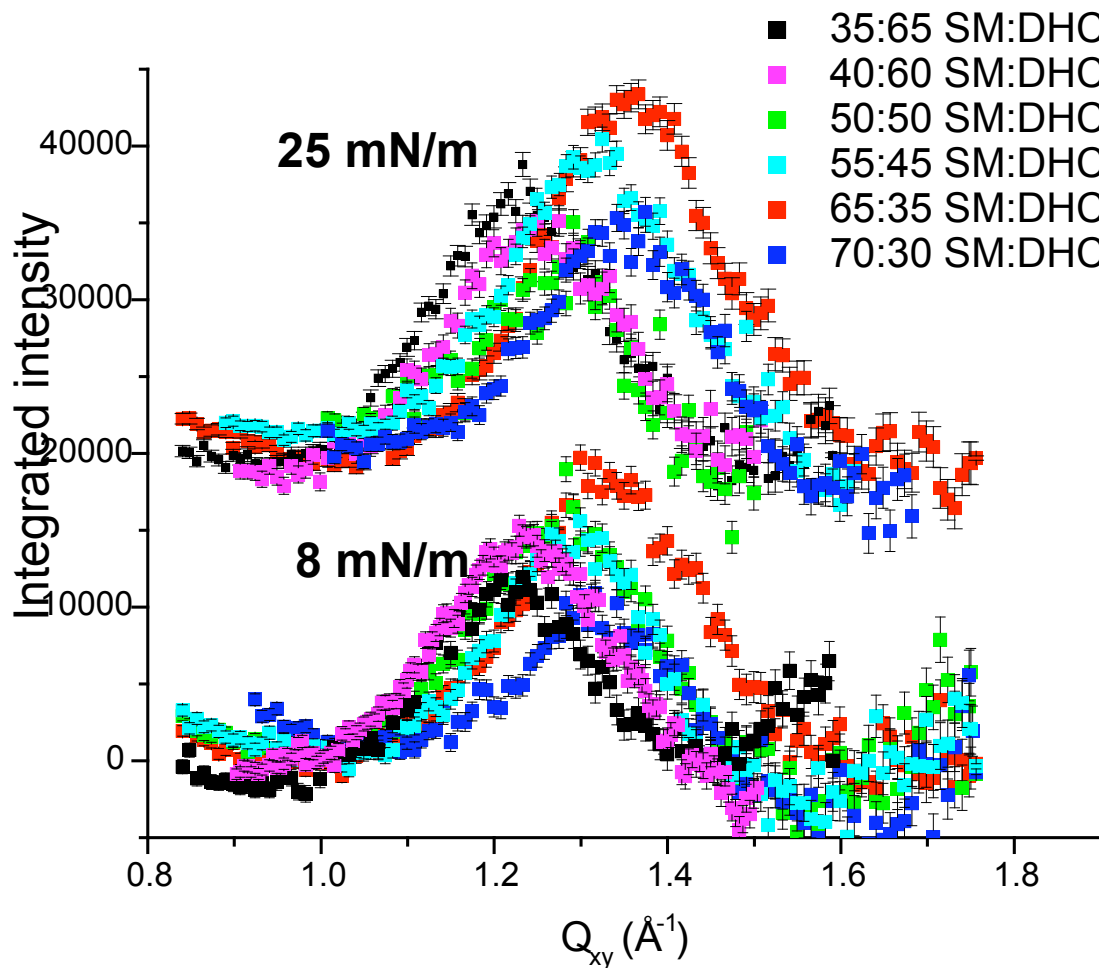
1:1 SM:DChol

Pressure	d spacing	coherence length	integrated intensity
25	4.86	24.39	1.0
30	4.85	23.61	1.0
35	4.84	23.38	1.1
40	4.80	22.25	1.3

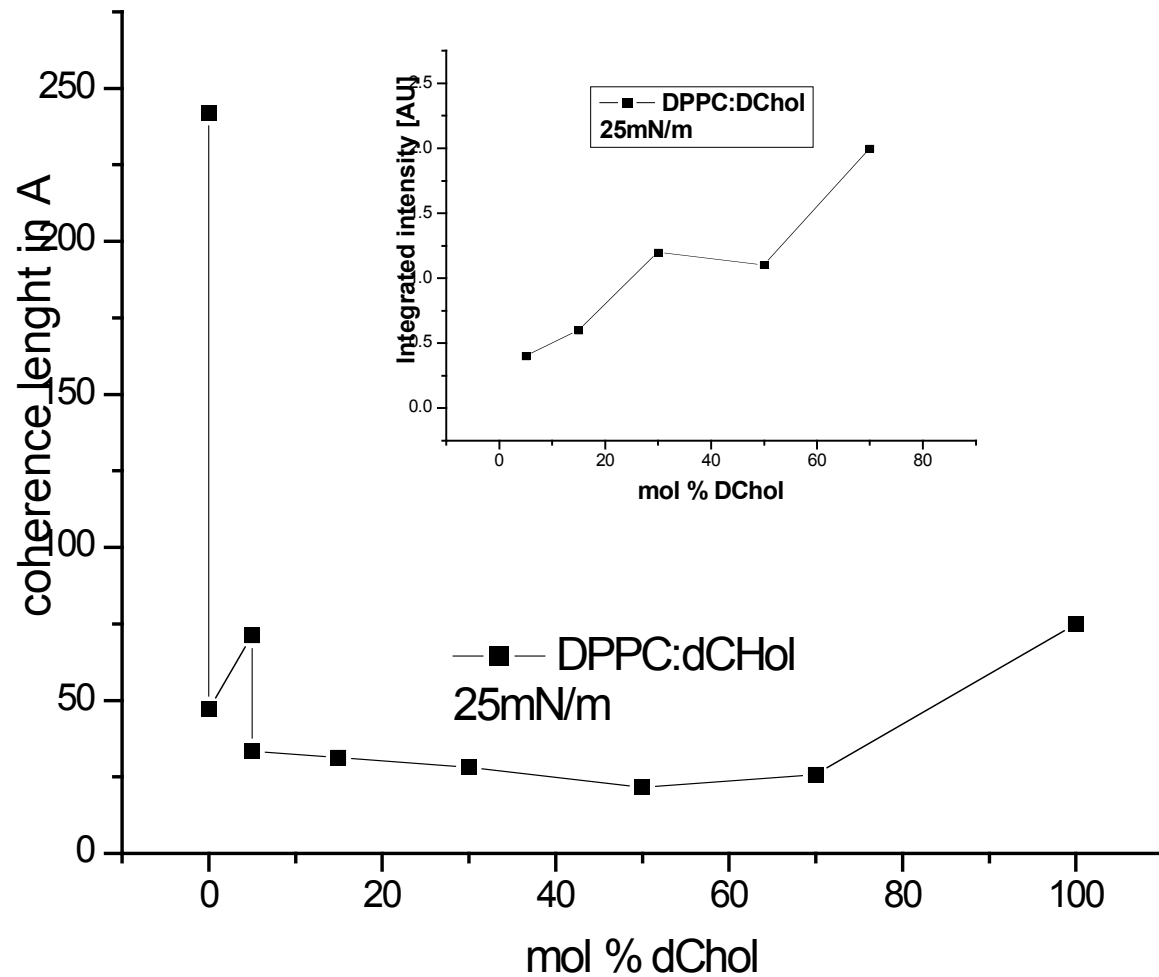
1:1:1 SM:DPPC:DChol

Pressure	d spacing	coherence length	integrated intensity
25	4.70	22.94	1.0
35	4.60	21.85	1.0
40	4.62	23.03	1.0

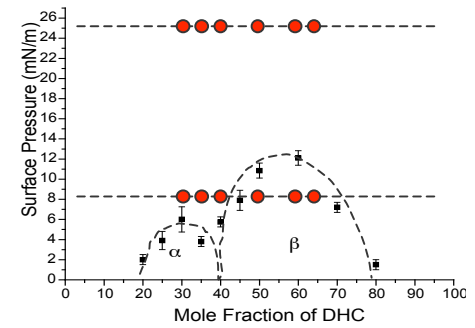
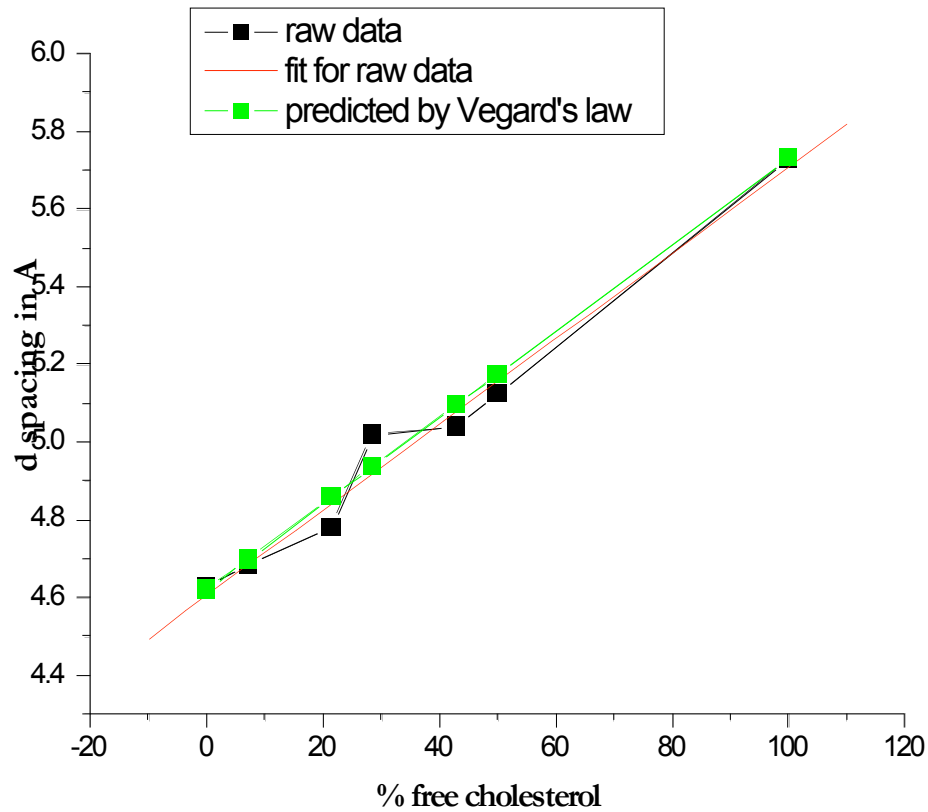
Structures Exist Below and Above Liquid-Liquid Immiscibility Line



Coherence Dependence on Chol Content in DPPC:Chol Systems

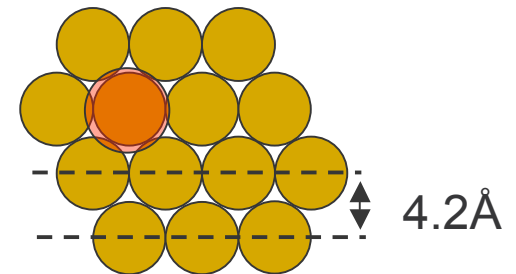
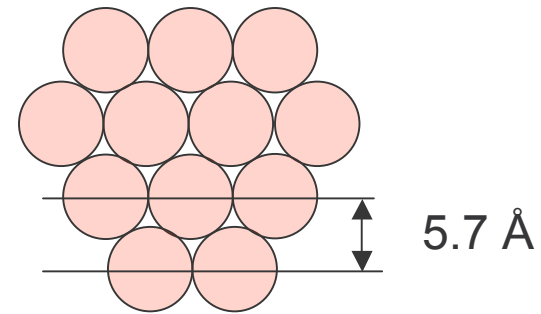
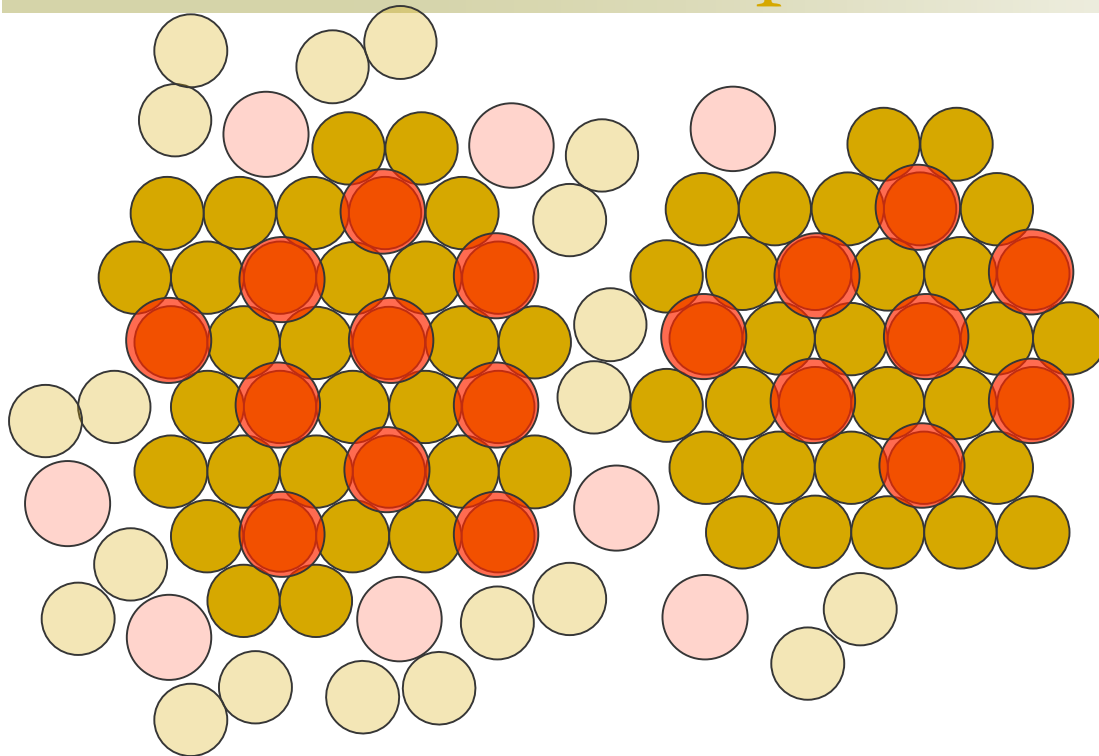






Vegard's Law



MR, JM, KK, KYCL, unpublished data (2007)

Proposed Model

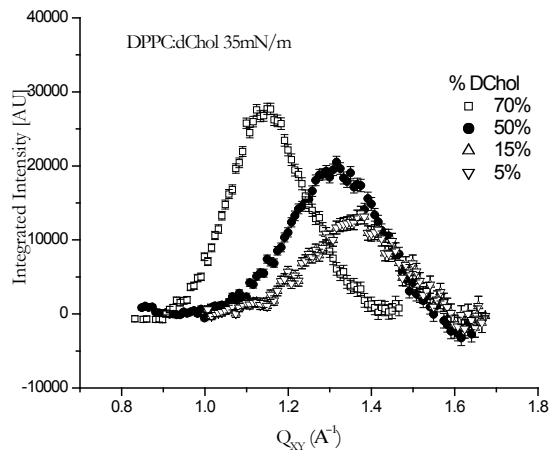


-  Uncomplexed phospholipid
-  Phospholipid in complex
-  Uncomplexed cholesterol
-  Cholesterol in complex

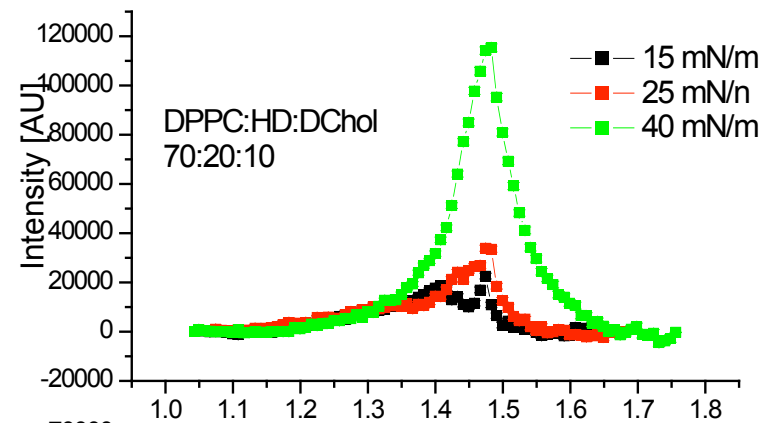
Dynamic Structure

Structural Signature of Displacement Effects

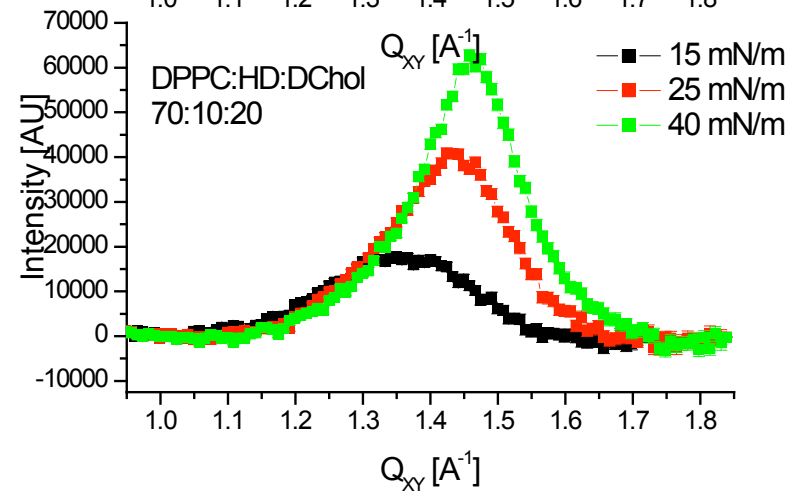
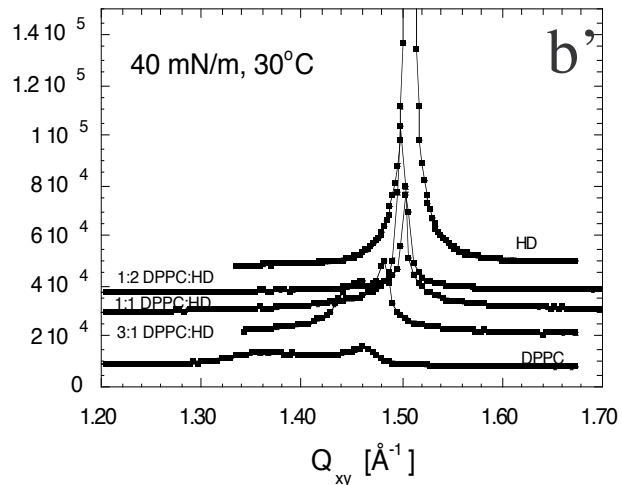
DPPC:DChol



DPPC:HD:DChol



DPPC:HD



Presence of DPPC:HD and DPPC:DChol peaks

DPPC:HD:DChol	Pressure (mN/m)	d-spacing (Å)	Coherence length (Å)	Integrated intensity (a.u)
100:0:0	15	-	-	-
	40	4.31, 4.57	150, 50	?
0:100:0	15	4.19	850	?
	40	4.17	620	?
0:0:100	25	5.71	63.1	1.7
	35	5.72	75.5	1.5
70:10:20	15	4.65	27.0	0.9
	25	4.37, 4.88	35.4, 43.3	1.6, 0.3
	40	4.30	33.3	3.5
70:20:10	15	4.27, 4.46, 4.80	311.6, 71.1, 35.4	0.1, 0.4, 0.4
	25	4.25, 4.36, 4.76	311.6, 81.4, 30.0	0.2, 0.4, 0.6
	40	4.25, 4.42	71.1, 56.8	2.8, 0.3
50:30:20	15	4.26, 4.42, 4.76	569, 78.4, 39.1	0.1, 0.5, 0.4
	25	4.27, 4.49, 4.77	68.6, 56.7, 33.0	0.9, 0.4, 0.5
	40	4.25, 4.52	56.7, 33.3	1.7, 0.9
50:40:10	15	4.25, 4.39	504.9, 130.9	0.5, 1.0
	25	4.25, 4.30, 4.62	1042.2, 114.7, 51.6	0.3, 2.0, 0.4
	40	4.19, 4.22	1042.2, 63.1	0.6, 3.0

Conclusions from X-ray Scattering Study

- Intermediate d-spacing provides evidence for cholesterol:phospholipid intermediate structures
- Reduced coherence lengths
 - => nanoscale domains
 - => dynamic structures
- Signatures of PC/HD and PC/DChol complexes observed in ternary mixtures

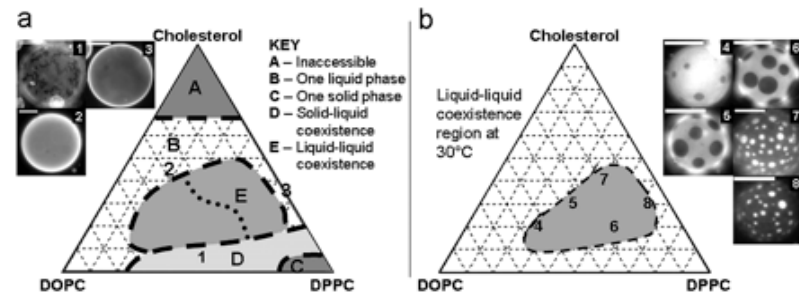
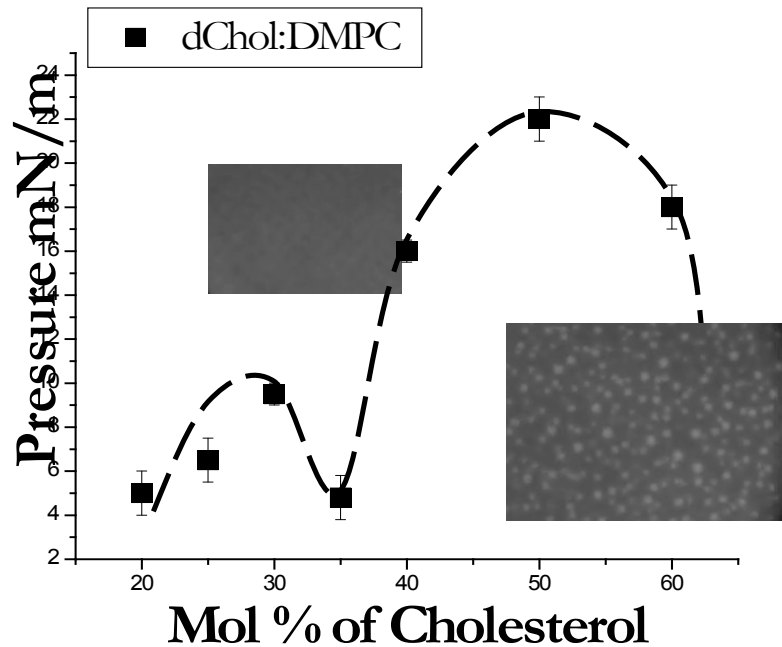
Four Different Approaches

- Phase diagrams
- Cholesterol desorption by cyclodextrin (CD)
- X-ray diffraction and x-ray reflectivity
- Effect of "inhibiting" and "promoting" sterols on cholesterol activity in RBC

Probing Free Cholesterol in RBC

- Cholesterol oxidase can oxidize cholesterol
 - causes the production of cholestenone only if there is excess (free) cholesterol available
 - Cholesterol oxidase depends on chemical activity of cholesterol
- The amount of cholestenone can be quantified by GC-FID

“Inhibiting” and “Promoting” Sterols

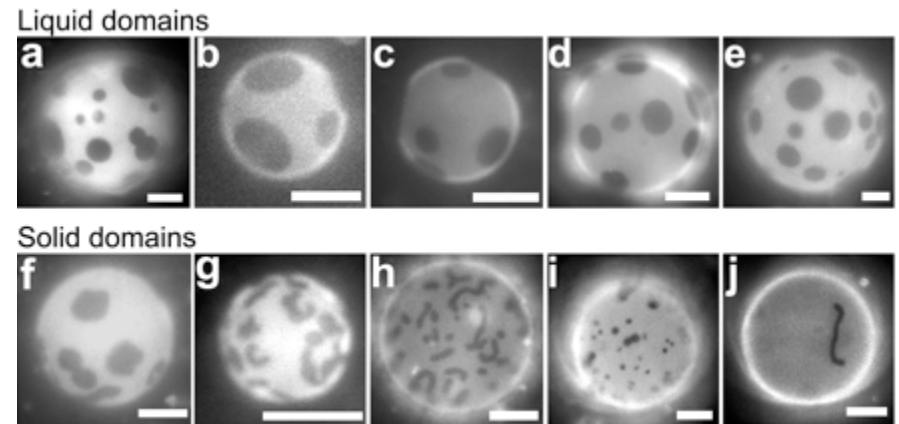
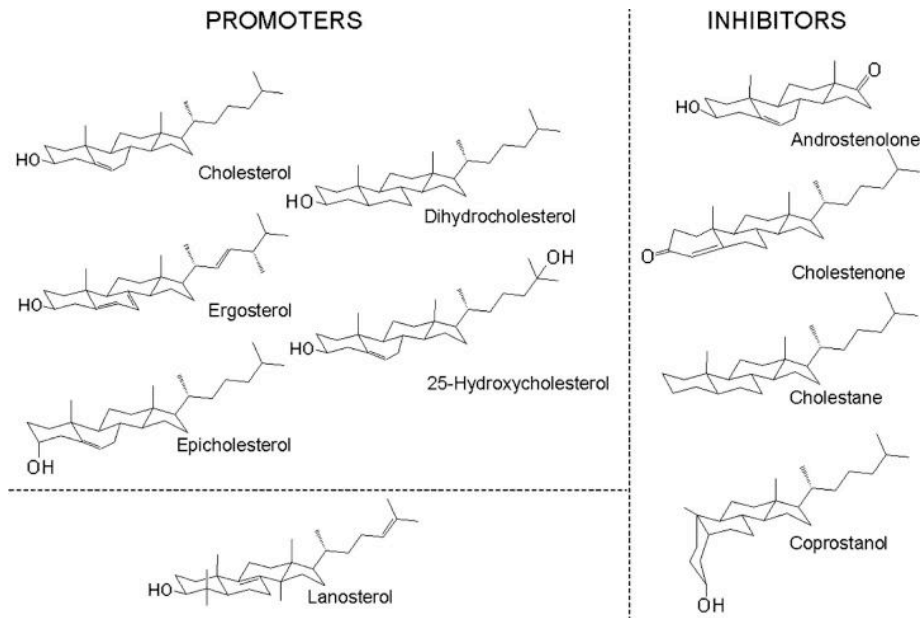


Observed phase diagram of micron-scale liquid immiscibility region in GUVs at 30°C.

Separation of Liquid Phases in Giant Vesicles of Ternary Mixtures of Phospholipids and Cholesterol
Sarah L. Veatch and Sarah L. Keller

Phase Behavior of Lipid Monolayers Containing DPPC and Cholesterol Analogs
Benjamin L. Stottrup and Sarah L. Keller

“Inhibiting” and “Promoting” Sterols

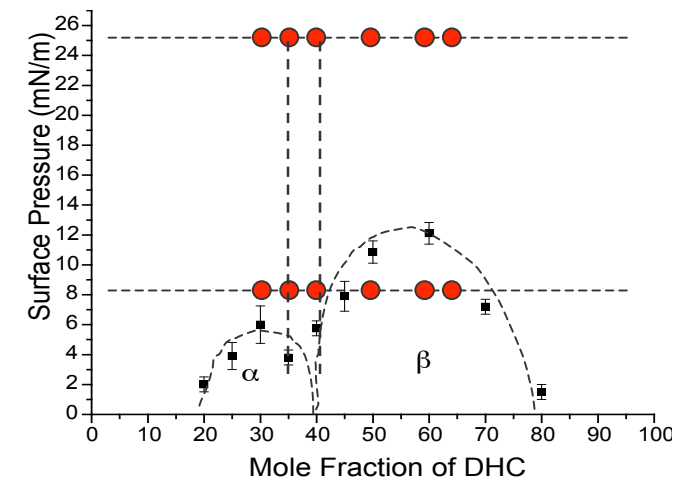
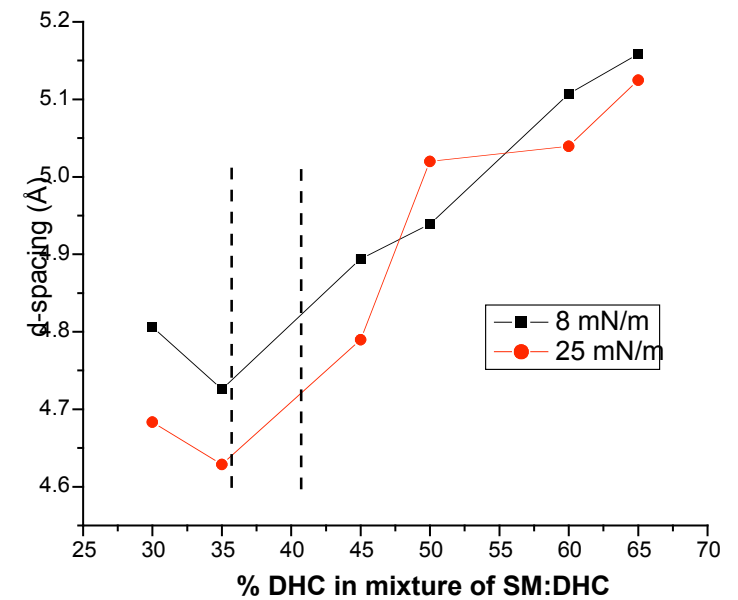
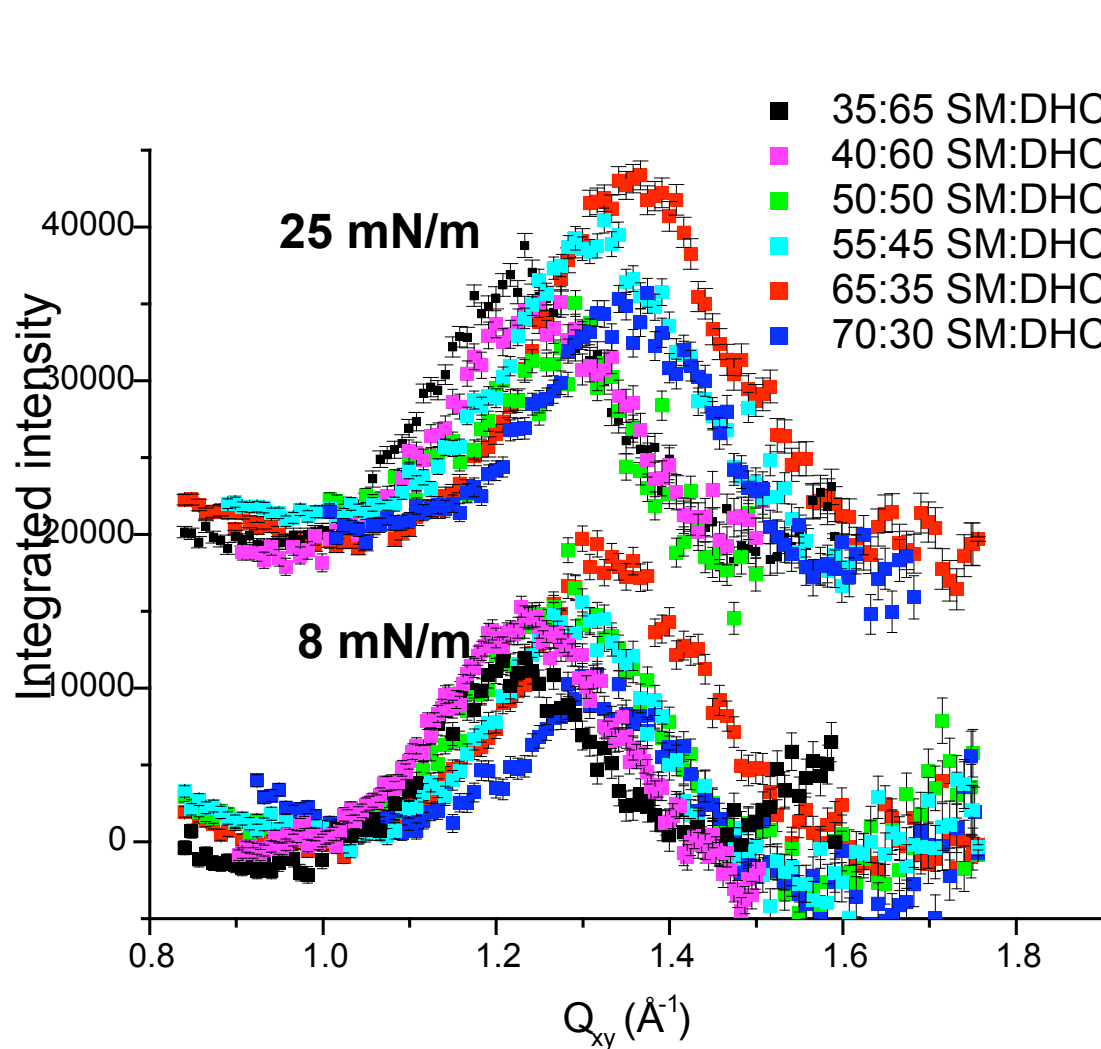


How does immiscibility correlates with active cholesterol?

■ Hypotheses:

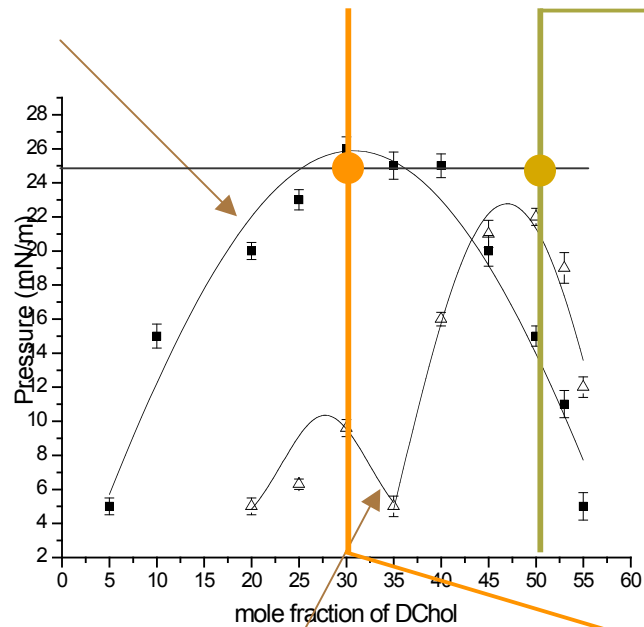
- Domains imply/require complex
- Lack of domains does not imply no complex
- Complex is necessary but not sufficient for domain phase separation
- Displacement of cholesterol by intercalators signifies complexation of intercalator with phospholipids, giving rise to free cholesterol
intercalators \Leftrightarrow cholesterol mimics

Structures Exist Below and Above Liquid-Liquid Immiscibility Line



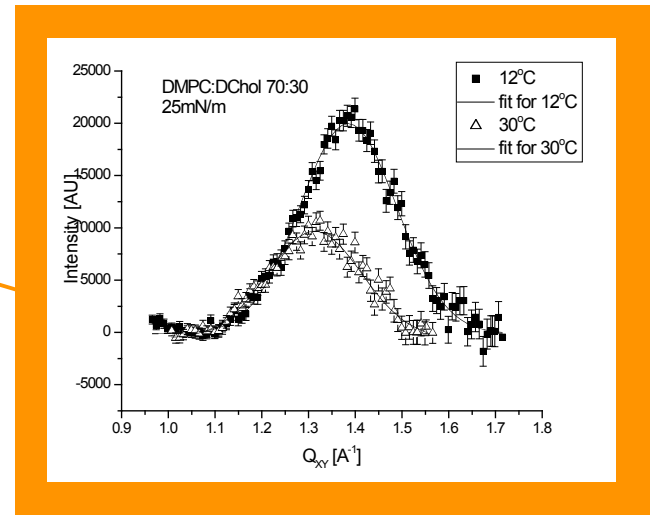
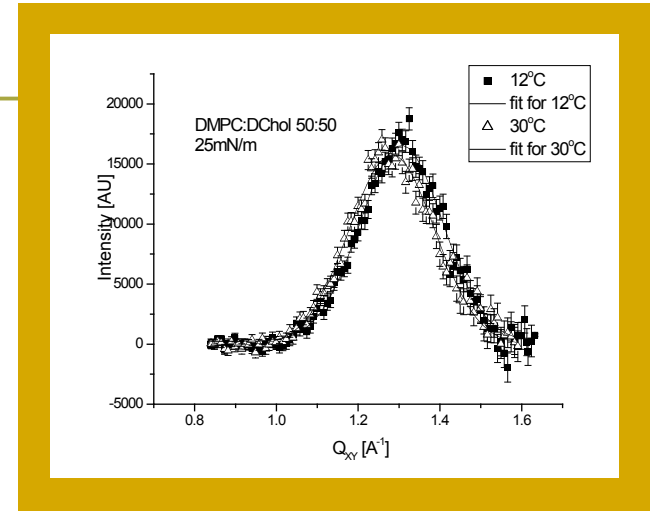
Temperature Dependence

$T = 30\text{ }^{\circ}\text{C}$



$T = 12\text{ }^{\circ}\text{C}$

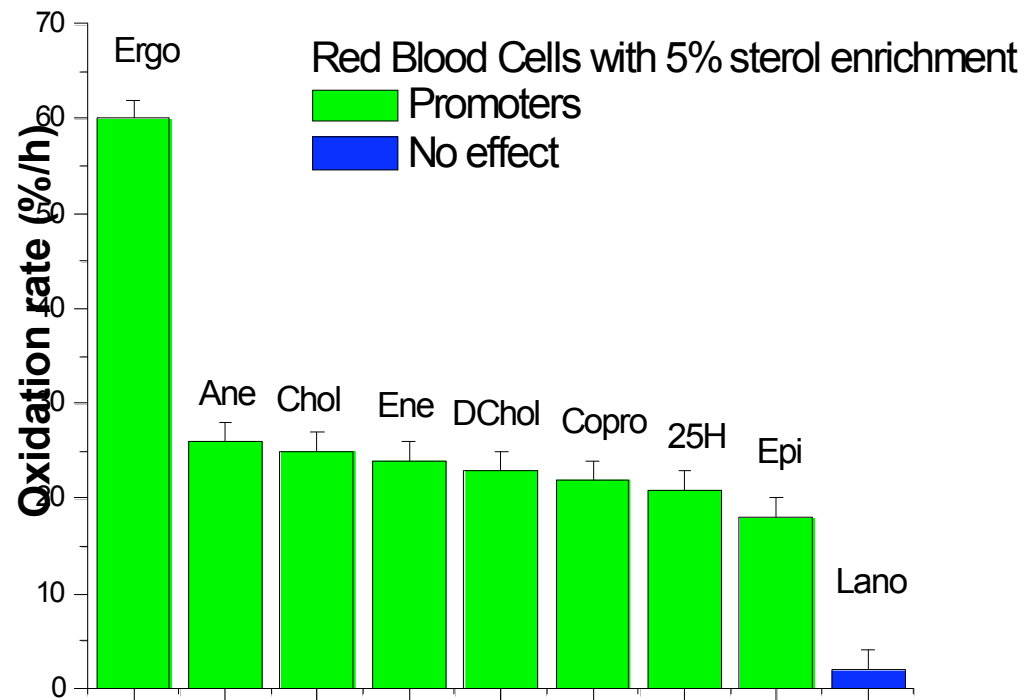
DMPC/DChol Binary Mixture



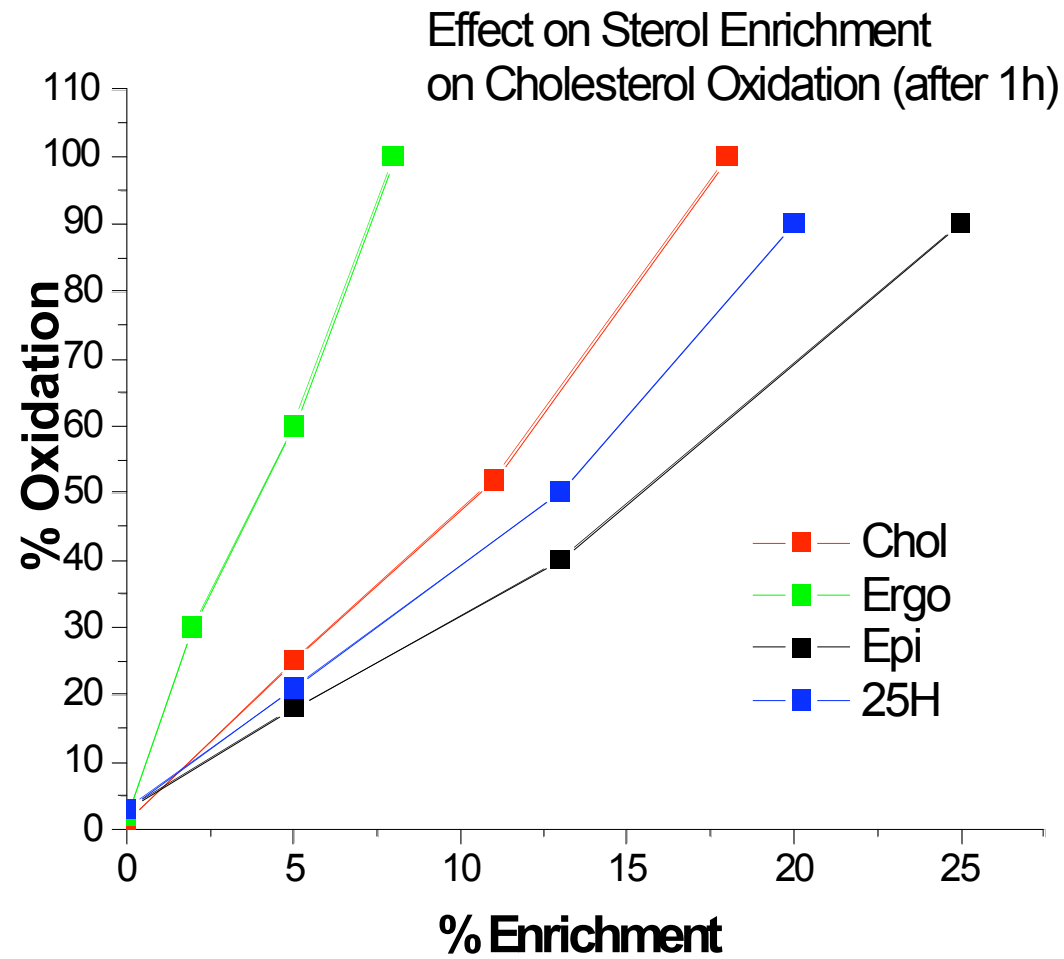
“Inhibiting” and “Promoting” Sterols

- If our hypotheses are correct:
 - should always observe active cholesterol with promoter sterols
 - could still observe active cholesterol even with inhibitor sterols
- What would disprove our hypotheses?
 - If we observe no active cholesterol in case of promoter sterols
 - This would mean that domains do not imply/depend upon complex formation

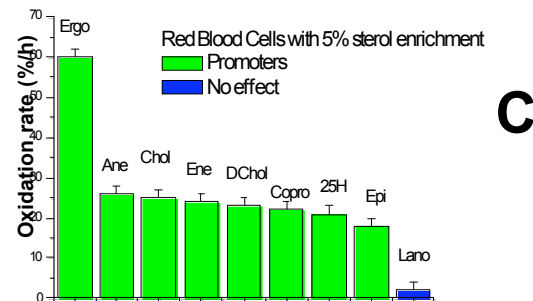
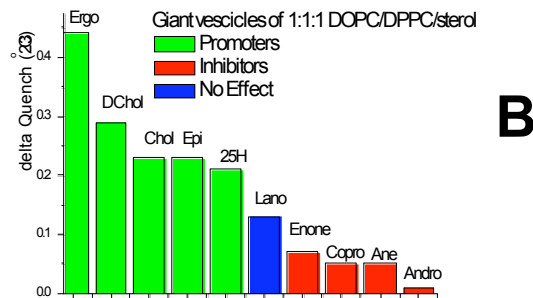
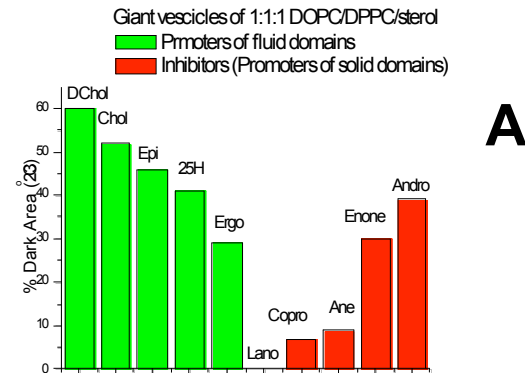
Testing for Cholestenone



Effect of Sterol Enrichment

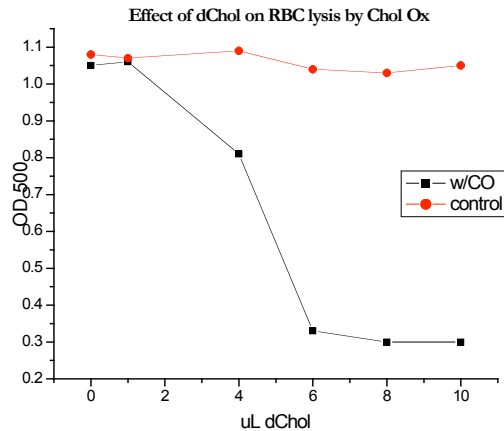


Displacement Observed for “Promoting” and “Inhibiting” Sterols

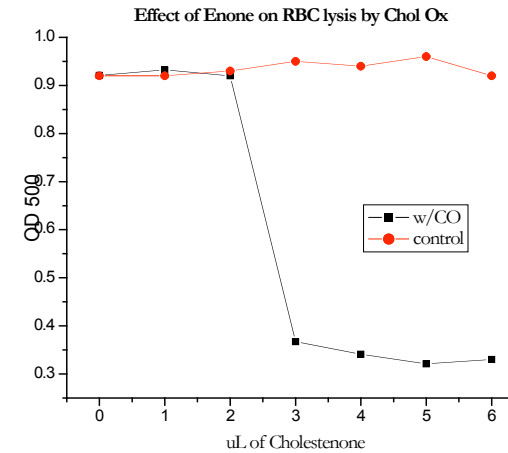


“Inhibiting” and “Promoting” Sterols Promoting Cell Lysis

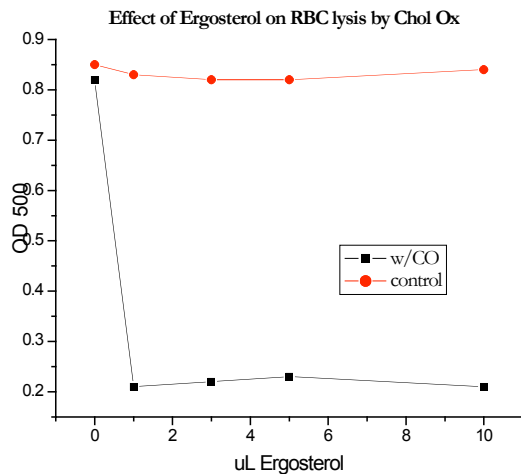
Dihydrocholesterol (P)



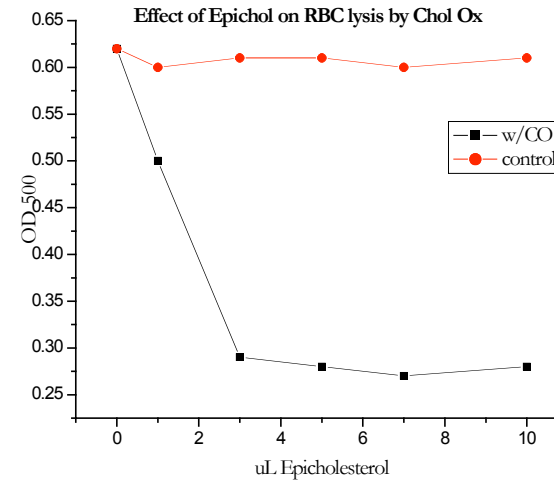
Cholestenone (I)



Ergosterol (P)



Epicholesterol (P)



Conclusions from RBC Study

- Tested all “promoting” sterols
 - All promoted cholestenone production and cell lysis by cholesterol oxidase
 - All displace cholesterol: produced free cholesterol available for cholesterol oxidase
- Cholesterol can exist in two states: bound to phospholipids and free

Conclusions

- 2 different states of cholesterol when present in lipid mixtures:
 - inactive (bound to lipid)
 - active (free)
- Production of active (free) cholesterol is possible even when actual cholesterol content is low but with alcohol present
- Alcohol displaces cholesterol from complexes and makes it available for cyclodextrin even with cholesterol content in the *alpha region*
- No cholesterol: quasi-long range order
- With cholesterol: very short-range order (coherence length $\sim 22 \text{ \AA}$); no pressure dependence
- Packing of phospholipid/cholesterol “complexes” depends on cholesterol content
- “Inhibiting” and “Promoting” sterols both can displace cholesterol

Acknowledgments

Canay

Maria

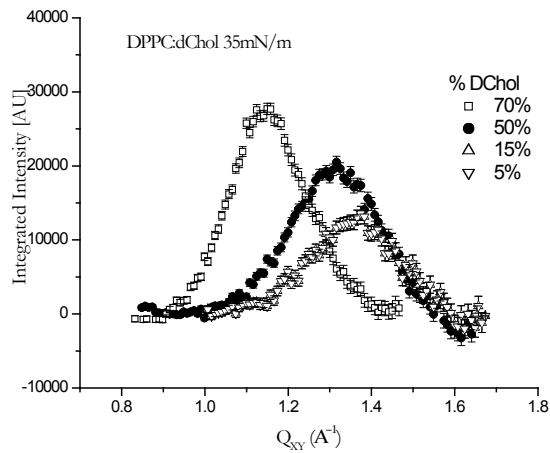
+ Chris (in UK)

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- Packard Foundation
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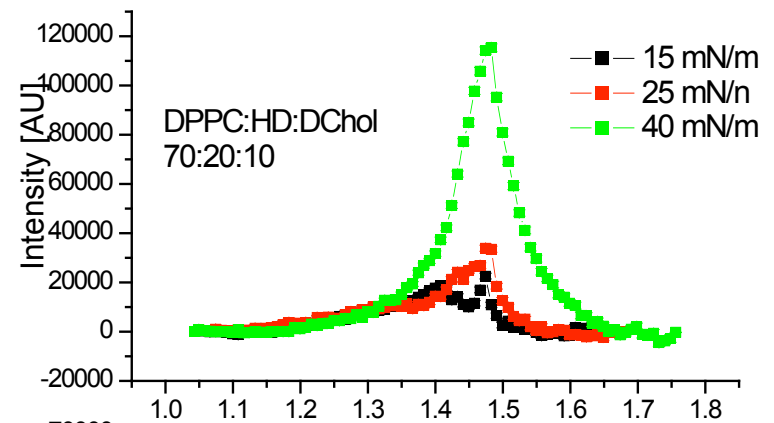


Structural Signature of Displacement Effects

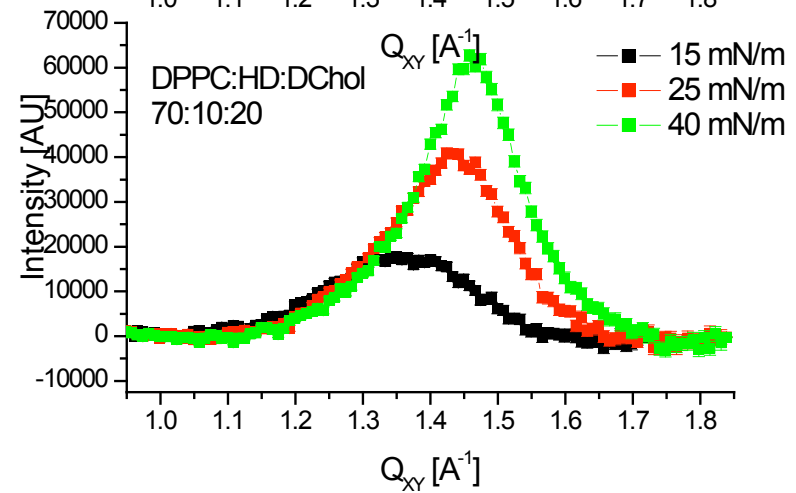
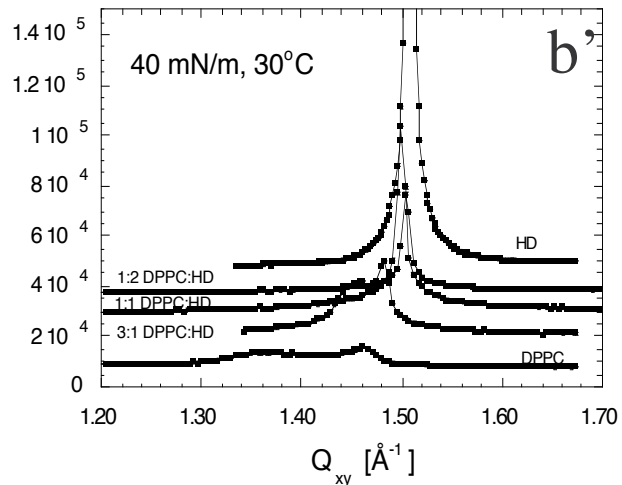
DPPC:DChol



DPPC:HD:DChol



DPPC:HD



Presence of DPPC:HD and DPPC:DChol peaks

DPPC:HD:DChol	Pressure (mN/m)	d-spacing (Å)	Coherence length (Å)	Integrated intensity (a.u)
100:0:0	15	-	-	-
	40	4.31, 4.57	150, 50	?
0:100:0	15	4.19	850	?
	40	4.17	620	?
0:0:100	25	5.71	63.1	1.7
	35	5.72	75.5	1.5
70:10:20	15	4.65	27.0	0.9
	25	4.37, 4.88	35.4, 43.3	1.6, 0.3
	40	4.30	33.3	3.5
70:20:10	15	4.27, 4.46, 4.80	311.6, 71.1, 35.4	0.1, 0.4, 0.4
	25	4.25, 4.36, 4.76	311.6, 81.4, 30.0	0.2, 0.4, 0.6
	40	4.25, 4.42	71.1, 56.8	2.8, 0.3
50:30:20	15	4.26, 4.42, 4.76	569, 78.4, 39.1	0.1, 0.5, 0.4
	25	4.27, 4.49, 4.77	68.6, 56.7, 33.0	0.9, 0.4, 0.5
	40	4.25, 4.52	56.7, 33.3	1.7, 0.9
50:40:10	15	4.25, 4.39	504.9, 130.9	0.5, 1.0
	25	4.25, 4.30, 4.62	1042.2, 114.7, 51.6	0.3, 2.0, 0.4
	40	4.19, 4.22	1042.2, 63.1	0.6, 3.0